Technical Manual

PowerPlex® Y System

INSTRUCTIONS FOR USE OF PRODUCTS DC6760 AND DC6761

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Part# TMD018

PowerPlex® Y System

All technical literature is available on the Internet at: www.promega.com/protocols/
Please visit the web site to verify that you are using the most current version of this Technical Manual.
Please contact Promega Technical Services if you have questions on use of this system.
E-mail: genetic@promega.com

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1. Description

STR (short tandem repeat) loci consist of short, repetitive sequence elements 3–7 base pairs in length (1–4). These repeats are well distributed throughout the human genome and are a rich source of highly polymorphic markers, which may be detected using the polymerase chain reaction (5–8). Alleles of STR loci are differentiated by the number of copies of the repeat sequence contained within the amplified region and are distinguished from one another using fluorescence detection following electrophoretic separation.

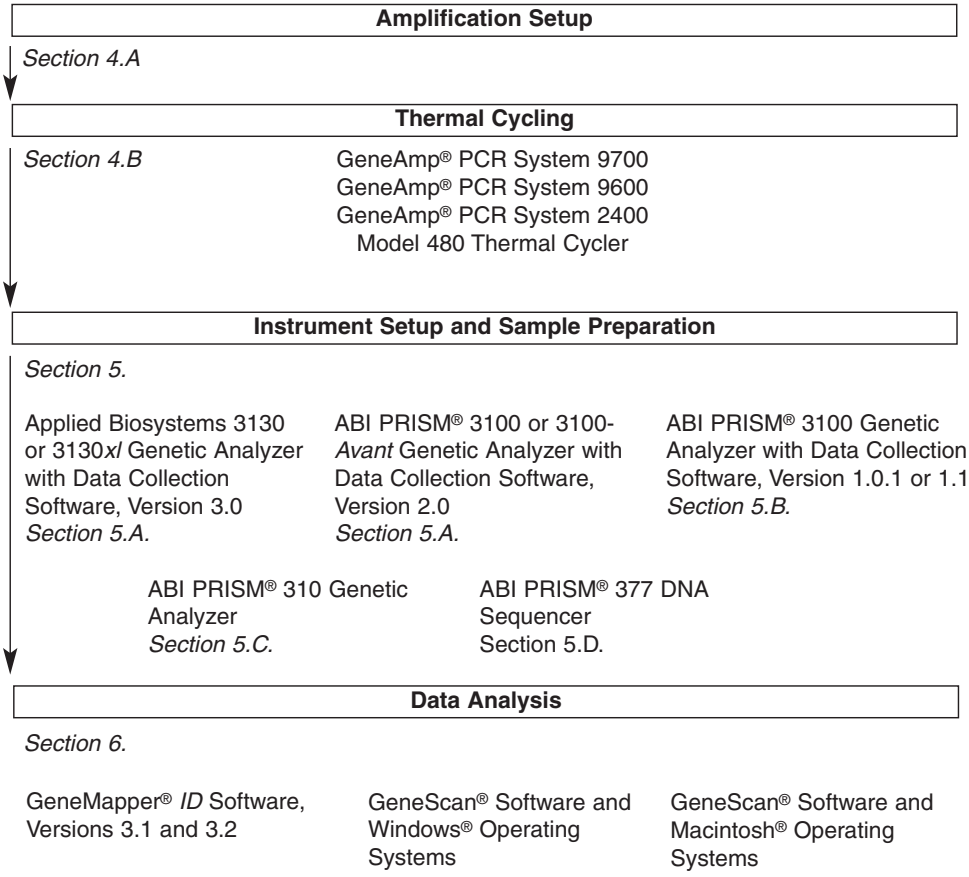
STR markers on the Y chromosome (Y-STR) have qualities that are distinct from autosomal markers and are useful for human identification (9–15). Y-STR markers are found on the nonrecombining region of the Y chromosome (NRY) and produce a haploid profile when amplified from male DNA. This quality simplifies male/female mixture interpretation by removing the female contribution from an amplification profile (16,17). Strict paternal inheritance of these markers makes them useful for paternity and kinship studies as well.

The PowerPlex® Y System[a,b] is used for human identification applications including forensic analysis, relationship testing and research use. This system allows co-amplification and three-color detection of twelve loci: DYS19, DYS385a/b, DYS389I/II, DYS390, DYS391, DYS392, DYS393, DYS437, DYS438 and DYS439 (18). One primer specific for each of the DYS389I/II, DYS391 and DYS439 loci is labeled with fluorescein (FL); one primer specific for each of the DYS385a/b, DYS390 and DYS393 loci is labeled with carboxy-tetramethyl-rhodamine (TMR); and one primer specific for each of the DYS19, DYS392, DYS437 and DYS438 loci is labeled with 6-carboxy-4´,5´-dichloro-2´,7´-dimethoxy-fluorescein (JOE). All twelve loci are amplified simultaneously in a single tube and analyzed in a single injection or gel lane.

The PowerPlex® Y System is compatible with the ABI PRISM® 310, 3100 and 3100-*Avant* Genetic Analyzers, Applied Biosystems 3130 and 3130*xl* Genetic Analyzers and ABI PRISM® 377 DNA Sequencer. The protocols presented in this manual were tested at Promega Corporation. Amplification and detection instrumentation may vary. You may need to optimize protocols including cycle number and injection conditions (or loading volume) for each laboratory instrument. In-house validation should be performed.

The PowerPlex® Y System provides all of the materials necessary to amplify Y-STR regions of purified human genomic DNA except for AmpliTaq Gold® DNA polymerase. This manual contains separate protocols for use of the PowerPlex® Y System with the Perkin-Elmer model 480 and GeneAmp® PCR system 9600, 9700 and 2400 thermal cyclers in addition to protocols to separate amplified products and detection of separated material (Figure 1). Protocols to operate the fluorescence-detection instruments should be obtained from the instrument manufacturer.

Information on other Promega fluorescent STR systems is available upon request from Promega or online at: **www.promega.com**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GeneScan® Software and Macintosh® Operating Systems

Figure 1. An overview of the PowerPlex® Y System protocol

2. Product Components and Storage Conditions

Product	Size	Cat.#
PowerPlex® Y System	50 reactions	DC6761

Not For Medical Diagnostic Use. Cat.# DC6761 contains sufficient reagents for 50 reactions of 25µl each. Includes:

Pre-amplification Components Box (Blue Label)

1 × 300µl	Gold ST★R 10X Buffer
1 × 125µl	PowerPlex® Y 10X Primer Pair Mix
25µl	2800M Control DNA, 10ng/µl

Post-amplification Components Box (Beige Label)

1 × 12.5µl	PowerPlex® Y Allelic Ladder Mix
1 × 150µl	Internal Lane Standard 600

Product	Size	Cat.#
PowerPlex® Y System	200 reactions	DC6760

Not For Medical Diagnostic Use. Cat.# DC6760 contains sufficient reagents for 200 reactions of 25µl each. Includes:

Pre-amplification Components Box (Blue Label)

2 × 300µl	Gold ST★R 10X Buffer
4 × 125µl	PowerPlex® Y 10X Primer Pair Mix
25µl	2800M Control DNA, 10ng/µl

Post-amplification Components Box (Beige Label)

4 × 12.5µl	PowerPlex® Y Allelic Ladder Mix
2 × 150µl	Internal Lane Standard 600

The PowerPlex® Y Allelic Ladder Mix is provided in a separate, sealed bag for shipping. This component should be moved to the post-amplification box after opening.

Storage Conditions: Store all components except the 2800M Control DNA at –30°C to –10°C in a nonfrost-free freezer. Store the 2800M Control DNA at 2 to 10°C. The PowerPlex® Y 10X Primer Pair Mix, PowerPlex® Y Allelic Ladder Mix and Internal Lane Standard (ILS) 600 are light-sensitive and must be stored in the dark. We strongly recommend that pre-amplification and post-amplification reagents be stored and used separately with different pipettes, tube racks, etc.

Available Separately

Product	Size	Cat.#
Blue Dextran Loading Solution*	3ml	DV4351
PowerTyper™ Macros (Release 2.0)**	1 CD-ROM	DG3470

*For Laboratory Use.
**Not For Medical Diagnostic Use.

The PowerTyper™ Macros (Release 2.0) for use with Genotyper® software are available from Promega. This CD-ROM contains the file "PowerTyperYMacroV2" for use with the PowerPlex® Y System. The macros can be also downloaded at: **www.promega.com/resources/tools/powertyper-macros/**

The proper panels and bins text files for use with GeneMapper® *ID* software can be obtained from the Promega web site at: **www.promega.com/resources/tools/genemapper-id-software-panels-and-bin-sets/**

Matrix standards are required for initial setup of the color separation matrix. The matrix standards are sold separately and are available for the ABI PRISM® 310 Genetic Analyzer and 377 DNA Sequencer (PowerPlex® Matrix Standards, 310) and the ABI PRISM® 3100 and 3100-*Avant* Genetic Analyzers and Applied Biosystems 3130 and 3130*xl* Genetic Analyzers (PowerPlex® Matrix Standards, 3100/3130). See Section 9.F for ordering information.

3. Before You Begin

3.A. Precautions

The application of PCR-based typing for forensic or paternity casework requires validation studies and quality-control measures that are not contained in this manual (19,20). The quality of the purified DNA, small changes in buffers, ionic strength, primer concentrations, choice of thermal cycler and thermal cycling conditions can affect PCR success. We suggest strict adherence to recommended procedures for amplification, as well as denaturing gel electrophoresis and fluorescence detection. Additional research and validation are required if any modifications are made to the recommended protocols.

PCR-based STR analysis is subject to contamination by very small amounts of human DNA. Extreme care should be taken to avoid cross-contamination when preparing sample DNA, handling primer pairs, assembling amplification reactions and analyzing amplification products. Reagents and materials used prior to amplification (Gold ST★R 10X Buffer, 2800M Control DNA and PowerPlex® Y 10X Primer Pair Mix) are provided in a separate box and should be stored separately from those used following amplification (PowerPlex® Y Allelic Ladder Mix and Internal Lane Standard 600). Always include a negative control reaction (i.e., no template) to detect reagent contamination. We highly recommend the use of gloves and aerosol-resistant pipette tips (e.g., ART® tips, Section 9.F).

3.A. Precautions (continued)

Some reagents used in the analysis of STR products are potentially hazardous and should be handled accordingly. Table 1 describes the potential hazards associated with such reagents.

Table 1. Hazardous Reagents.

Reagents for ABI PRISM® 310, 3100 and 3100-*Avant* Genetic Analyzers and Applied Biosystems 3130 and 3130*xl* Genetic Analyzers	Hazard
formamide	irritant, teratogen

Reagents for ABI PRISM® 377 DNA Sequencer	Hazard
acrylamide (Long Ranger® Gel Solution)	suspected carcinogen, toxic
ammonium persulfate	oxidizer, corrosive
formamide (contained in the Blue Dextran Loading Solution)	irritant, teratogen
TEMED	corrosive, flammable
urea	irritant

3.B. Matrix Standardization or Spectral Calibration

Proper generation of a matrix file is critical to evaluate multicolor systems with the ABI PRISM® 310, 3100 and 3100-*Avant* Genetic Analyzers and Applied Biosystems 3130 and 3130*xl* Genetic Analyzers. A matrix must be generated for each individual instrument.

The PowerPlex® Matrix Standards, 310 (Cat.# DG4640), is required for matrix standardization for the ABI PRISM® 310 Genetic Analyzer and ABI PRISM® 377 DNA Sequencer. The PowerPlex® Matrix Standards, 3100/3130 (Cat.# DG4650), cannot be used to generate a matrix on the ABI PRISM® 310 Genetic Analyzer.

The PowerPlex® Matrix Standards, 3100/3130 (Cat.# DG4650), is required for spectral calibration on the ABI PRISM® 3100 and 3100-*Avant* Genetic Analyzers and Applied Biosystems 3130 and 3130*xl* Genetic Analyzers. The PowerPlex® Matrix Standards, 310 (Cat.# DG4640), cannot be used to generate a matrix on these instruments.

For protocols and additional information on matrix standardization, see the *PowerPlex® Matrix Standards, 310, Technical Bulletin* #TBD021, which is supplied with Cat.# DG4640. For protocols and additional information on spectral calibration, see the *PowerPlex® Matrix Standards, 3100/3130, Technical Bulletin* #TBD022, which is supplied with Cat.# DG4650. These manuals are available online at: **www.promega.com/protocols/**

4. Protocols for DNA Amplification Using the PowerPlex® Y System

Materials to Be Supplied by the User

- model 480 or GeneAmp® PCR System 9600, 9700 or 2400 thermal cycler (Applied Biosystems)
- microcentrifuge
- 0.5ml or 0.2ml thin-walled reaction tubes or MicroAmp® optical 96-well reaction plate (Applied Biosystems)
- aerosol-resistant pipette tips (see Section 9.F)
- AmpliTaq Gold® DNA polymerase (Applied Biosystems)
- Nuclease-Free Water (Cat.# P1193)
- Mineral Oil (Cat.# DY1151, for use with the model 480 thermal cycler)

We routinely amplify 0.5–1ng of male template DNA in a 25µl reaction volume using the protocols detailed below. Expect to see high peak heights at the smaller loci and relatively lower peak heights at larger loci if more than the recommended amount of male template DNA is used. Reduce the amount of male template DNA or the number of cycles to correct this.

The PowerPlex® Y System is optimized for the GeneAmp® PCR System 9700 thermal cycler. Amplification protocols for the GeneAmp® PCR Systems 9600 and 2400 thermal cyclers and Perkin-Elmer model 480 thermal cycler also are provided.

A mixture of male and female DNA will often necessitate the use of more than 1ng of total DNA (male and female DNA combined). This system has been designed to amplify a male-derived haplotype even in the presence of female DNA. The range of total input DNA and the ratio of male to female DNA that produces acceptable results should be validated in your laboratory. Amplification and analysis of the Amelogenin locus prior to multiplex analysis may provide some information as to the ratio of male to female DNA. Amelogenin is available as a monoplex amplification with a ladder labeled with either fluorescein (Cat.# DC5171) or TMR (Cat.# DC6171).

4.A. Amplification Setup

The use of gloves and aerosol-resistant pipette tips is highly recommended to prevent cross-contamination. Keep all pre-amplification and post-amplification reagents in separate rooms. Prepare amplification reactions in a room dedicated for reaction setup. Use equipment and supplies dedicated for amplification setup.

Meticulous care must be taken to ensure successful amplification. A guide to amplification troubleshooting is provided in Section 7.A.

The concentration of 2800M Control DNA was determined by measuring absorbance at 260nm. Quantification of this control DNA by other methods, such as qPCR, may result in a different value. Prepare a fresh DNA dilution for each set of amplifications. Do not store diluted DNA (e.g., 0.25ng/µl or less).

4.A. Amplification Setup (continued)

1. Thaw the Gold ST★R 10X Buffer and PowerPlex® Y 10X Primer Pair Mix.

   Notes:

   1. Mix reagents by vortexing each tube for 15 seconds before each use. Do not centrifuge the 10X Primer Pair Mix after vortexing, as this may cause the primers to be concentrated at the bottom of the tube.
   2. A precipitate may form in the Gold ST★R 10X Buffer. If this occurs, warm the solution briefly at 37°C, then vortex until the precipitate is in solution.

2. Determine the number of reactions to be set up. This should include positive and negative control reactions. Add 1 or 2 reactions to this number to compensate for pipetting error. While this approach does consume a small amount of each reagent, it ensures that you will have enough PCR amplification mix for all samples. It also ensures that each reaction contains the same PCR amplification mix.

3. Place one clean, 0.2ml or 0.5ml reaction tube for each reaction into a rack, and label appropriately. Alternatively, use a MicroAmp® plate, and label appropriately.

   Note: If using the GeneAmp® PCR System 9600, 9700 or 2400 thermal cyclers, use 0.2ml MicroAmp® 8-strip reaction tubes or MicroAmp® plate. For the Perkin-Elmer model 480, we recommend standard 0.5ml GeneAmp® thin-walled reaction tubes.

4. Add the final volume of each reagent listed in Table 2 into a sterile, tube. Mix gently.

   Table 2 shows the component volumes per reaction. A worksheet to calculate the required amount of each PCR amplification mix component is provided in Section 9.D (Table 6).

   Amplification of >1ng of male DNA template results in an imbalance in peak heights from locus to locus. The smaller loci show greater amplification yield than the larger loci. Reducing the number of cycles in the amplification program by 2 to 4 cycles (i.e., 10/20 or 10/18 cycling) can improve locus-to-locus balance.

5. Pipet PCR amplification mix into each reaction tube.

6. Pipet the template DNA (0.5–1ng) for each sample into the respective tube containing PCR amplification mix.

7. For the positive amplification control, vortex the tube of 2800M Control DNA, then dilute an aliquot to 0.5ng in the desired template DNA volume. Pipet 0.5ng of the diluted DNA into a reaction tube containing PCR amplification mix.

   Note: To store diluted 2800M Control DNA, dilute the DNA to 0.5ng/µl in TE^{-4} buffer with 20µg/ml glycogen and store at 4°C. Do not store dilutions performed in water.

Table 2. PCR Amplification Mix for the PowerPlex® Y System.

PCR Amplification Mix Component[1]	Volume Per Reaction
nuclease-free water	to a final volume of 25.0µl
Gold ST★R 10X Buffer	2.5µl
PowerPlex® Y 10X Primer Pair Mix	2.5µl
AmpliTaq Gold® DNA polymerase[2]	0.55µl (2.75u)
template DNA (up to 1ng)[3]	up to 19.45µl
total reaction volume	**25µl**

[1]Add nuclease-free water to the PCR amplification mix first, then add Gold ST★R 10X Buffer, PowerPlex® Y 10X Primer Pair Mix and AmpliTaq Gold® DNA polymerase. The template DNA will be added at Step 6.

[2]Assumes the AmpliTaq Gold® DNA polymerase is at 5u/µl. If the enzyme concentration is different, the volume of enzyme must be adjusted accordingly.

[3]Store DNA templates in nuclease-free water or TE^{-4} buffer (10mM Tris-HCl [pH 8.0], 0.1mM EDTA). If the DNA template is stored in TE buffer that is not pH 8.0 or contains a higher EDTA concentration, the volume of DNA added should not exceed 20% of the final reaction volume. Amplification efficiency and quality can be greatly altered by changes in pH (due to added Tris-HCl), available magnesium concentration (due to chelation by EDTA) or other PCR inhibitors, which may be present at low concentrations depending on the source of the template DNA and the extraction procedure used.

8. For the negative amplification control, pipet nuclease-free water or TE^{-4} buffer instead of template DNA into a reaction tube containing PCR amplification mix.

9. If using the GeneAmp® PCR System 9600, 9700 or 2400 thermal cycler and MicroAmp® reaction tubes or plates, no addition of mineral oil to the reaction tubes is required. However, if using the model 480 thermal cycler and GeneAmp® reaction tubes, add one drop of mineral oil to each tube before closing.

   Note: Allow the mineral oil to flow down the side of the tube and form an overlay to limit sample loss or cross-contamination due to splattering.

10. **Optional:** Briefly centrifuge the tubes to bring contents to the bottom and remove any air bubbles.

4.B. Amplification Thermal Cycling

This manual contains protocols for use of the PowerPlex® Y System with the Perkin-Elmer model 480 and GeneAmp® PCR system 9600, 9700 and 2400 thermal cyclers. For information on other thermal cyclers, please contact Promega Technical Services by e-mail: **genetic@promega.com**

Amplification and detection instrumentation may vary. You may need to optimize protocols including cycle number and injection conditions (or loading volume) for each laboratory instrument. Testing at Promega Corporation shows that 10/22 cycles work well for 0.5–1ng of purified DNA templates. For higher amounts of input DNA or to decrease sensitivity, fewer cycles, such as 10/16, 10/18 or 10/20, should be evaluated. In-house validation should be performed.

1. Place the tubes or MicroAmp® plate in a thermal cycler.
2. Select and run a recommended protocol. The preferred protocols for use with the GeneAmp® PCR System 9600, 9700 and 2400 thermal cyclers and Perkin-Elmer model 480 thermal cycler are provided below.
3. After completion of the thermal cycling protocol, store the amplified samples at –20°C in a light-protected box.

   Note: Long-term storage of amplified samples at 4°C or higher may produce degradation products.

Protocol for the GeneAmp® PCR System 9700 Thermal Cycler[1]	Protocol for the GeneAmp® PCR System 2400 Thermal Cycler
95°C for 11 minutes, then:	95°C for 11 minutes, then:
96°C for 1 minute, then:	96°C for 1 minute, then:
ramp 100% to 94°C for 30 seconds ramp 29% to 60°C for 30 seconds ramp 23% to 70°C for 45 seconds for 10 cycles, then:	ramp 100% to 94°C for 30 seconds ramp 100% to 60°C for 30 seconds ramp 23% to 70°C for 45 seconds for 10 cycles, then:
ramp 100% to 90°C for 30 seconds ramp 29% to 58°C for 30 seconds ramp 23% to 70°C for 45 seconds for 22 cycles, then:	ramp 100% to 90°C for 30 seconds ramp 100% to 58°C for 30 seconds ramp 23% to 70°C for 45 seconds for 22 cycles, then:
60°C for 30 minutes	60°C for 30 minutes
4°C soak	4°C soak
Protocol for the GeneAmp® PCR System 9600 Thermal Cycler	**Protocol for the Perkin-Elmer Model 480 Thermal Cycler**
95°C for 11 minutes, then:	95°C for 11 minutes, then:
96°C for 1 minute, then:	96°C for 2 minutes, then:
94°C for 30 seconds ramp 68 seconds to 60°C (hold for 30 seconds) ramp 50 seconds to 70°C (hold for 45 seconds) for 10 cycles, then:	94°C for 1 minute 60°C for 1 minute 70°C for 1.5 minutes for 10 cycles, then:
90°C for 30 seconds ramp 60 seconds to 58°C (hold for 30 seconds) ramp 50 seconds to 70°C (hold for 45 seconds) for 22 cycles, then:	90°C for 1 minute 58°C for 1 minute 70°C for 1.5 minutes for 22 cycles, then:
60°C for 30 minutes	60°C for 30 minutes
4°C soak	4°C soak

[1]When using the GeneAmp® PCR System 9700 thermal cycler, the ramp rates indicated in the cycling program must be set, and the program must be run in 9600 ramp mode.

The ramp rates are set in the Ramp Rate Modification screen. While viewing the cycling program, navigate to the Ramp Rate Modification screen by selecting "More", then "Modify". On the Ramp Rate Modification screen the default rates for each step are 100%. The rate under each hold step is the rate at which the temperature will change to that hold temperature. Figure 2 shows the ramp rates for the GeneAmp® PCR System 9700 thermal cycler.

The ramp mode is set after “start” has been selected for the thermal cycling run. A Select Method Options screen appears. Select 9600 ramp mode, and enter the reaction volume.

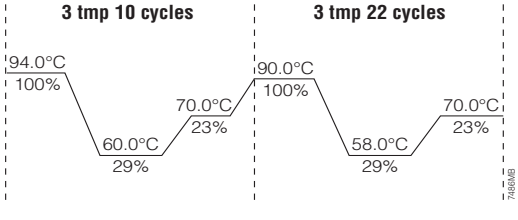

Figure 2. The ramp rates for the GeneAmp® PCR System 9700 thermal cycler.

5. Instrument Setup and Sample Preparation

5.A. Detection of Amplified Fragments Using the ABI PRISM® 3100 or 3100-*Avant* Genetic Analyzer with Data Collection Software, Version 2.0, or the Applied Biosystems 3130 or 3130*xl* Genetic Analyzer with Data Collection Software, Version 3.0

Materials to Be Supplied by the User

- 95°C dry heating block, water bath or thermal cycler
- crushed ice or ice-water bath
- aerosol-resistant pipette tips
- 3100 or 3130 capillary array, 36cm
- performance optimized polymer 4 (POP-4™) for the 3100 or 3130
- 10X genetic analyzer buffer with EDTA
- MicroAmp® optical 96-well plate and septa
- Hi-Di™ formamide (Applied Biosystems Cat.# 4311320)
- PowerPlex® Matrix Standards, 3100/3130 (Cat.# DG4650)

The quality of formamide is critical. Use Hi-Di™ formamide. Freeze formamide in aliquots at –20°C. Multiple freeze-thaw cycles or long-term storage at 4°C may cause breakdown of formamide. Poor-quality formamide may contain ions that compete with DNA during injection, which results in lower peak heights and reduced sensitivity. A longer injection time may not increase the signal.

Formamide is an irritant and a teratogen; avoid inhalation and contact with skin. Read the warning label, and take appropriate precautions when handling this substance. Always wear gloves and safety glasses when working with formamide.

Sample Preparation

1. Prepare a loading cocktail by combining and mixing Internal Lane Standard 600 and Hi-Di™ formamide as follows:

   [(0.5µl ILS 600) × (# injections)] + [(9.5µl Hi-Di™ formamide) × (# injections)]

   Note: The volume of internal lane standard used in the loading cocktail can be increased or decreased to adjust the intensity of the size standard peaks. The optimal peak height for the 100-base fragment of the internal lane standard is 500–1,000RFU. If peak heights are too low, we recommend altering the formamide/internal lane standard mix to contain 1.0µl of ILS 600 and 9.0µl of Hi-Di™ formamide. If peak heights are too high, we recommend altering the loading cocktail to contain 0.25µl of ILS 600 and 9.75µl of formamide.

2. Vortex for 10–15 seconds to mix.

3. Pipet 10µl of formamide/internal lane standard mix into each well.

4. Add 1µl of amplified sample (or 1µl of PowerPlex® Y Allelic Ladder Mix). Cover wells with appropriate septa.

   Note: Instrument detection limits vary; therefore, injection time, injection voltage or the amount of product mixed with loading cocktail may need to be adjusted. Use the Module Manager in the data collection software to modify the injection time or voltage in the run module. If peak heights are higher than desired, samples can be diluted in Gold ST★R 1X Buffer before mixing with loading cocktail. The use of too much template DNA may result in uneven allele peak heights across loci. For best results, use less DNA template in the amplification reactions or reduce the number of cycles in the amplification program by 2–4 cycles to achieve the desired signal intensity.

5. Centrifuge plate briefly to remove air bubbles from the wells.

6. Denature samples at 95°C for 3 minutes, then immediately chill on crushed ice or in an ice-water bath for 3 minutes. Denature samples just prior to loading the instrument.

Instrument Preparation

Refer to the instrument users' manual for instructions on cleaning, installing the capillary array, performing a spatial calibration and adding polymer.

Analyze samples as described in the user's manual for the ABI PRISM® 3100 or 3100-*Avant* Genetic Analyzer with Data Collection Software, Version 2.0, and the Applied Biosystems 3130 or 3130*xl* Genetic Analyzer with the following exceptions.

1. In the Module Manager, select "New". Select "Regular" in the Type drop-down list, and select "HIDFragmentAnalysis36_POP4" in the Template drop-down list. Confirm that the injection time is 5 seconds and the injection voltage is 3kV. Lengthen the run time to 2,000 seconds. Give a new name to your run module, and select "OK".

   Note: Instrument sensitivities can vary. The injection time and voltage may be adjusted in the Module Manager. A suggested range for the injection time is 3–22 seconds and for the injection voltage is 1–3kV.

2. In the Protocol Manager, select "New". Type a name for your protocol. Select "Regular" in the Type drop-down list, and select the run module you created in the previous step in the Run Module drop-down list. Lastly, select "F" in the Dye-Set drop-down list. Select "OK".

3. In the Plate Manager, create a new plate record as described in the instrument user's manual. In the dialog box that appears, select "GeneMapper—Generic" in the Application drop-down list, and select the appropriate plate type (96-well). Add entries in the owner and operator windows, and select "OK".

   Note: If autoanalysis of sample data is desired, refer to the instrument user's manual for instructions.

5.A. Detection of Amplified Fragments Using the ABI PRISM® 3100 or 3100-*Avant* Genetic Analyzer with Data Collection Software, Version 2.0, or the Applied Biosystems 3130 or 3130*xl* Genetic Analyzer with Data Collection Software, Version 3.0 (continued)

4. In the GeneMapper® plate record, enter sample names in the appropriate cells. Scroll to the right. In the Results Group 1 column, select the desired results group. In the Instrument Protocol 1 column, select the protocol you created in Step 2. Be sure this information is present for each row that contains a sample name. Select "OK".

   Note: To create a new Results Group, select "New" in the drop-down menu in the Results Group column. Select the General tab, and enter a name. Select the Analysis tab, and select "GeneMapper—Generic" in the Analysis type drop-down list.

5. Place samples in the instrument, and close the instrument doors.

6. In the spectral viewer, confirm that dye set F is active, and set the correct active calibration for dye set F.

7. In the run scheduler, locate the plate record that you just created in Steps 3 and 4, and click once on the name to highlight it.

8. Once the plate record is highlighted, click the plate graphic that corresponds to the plate on the autosampler that contains your amplified samples.

9. When the plate record is linked to the plate, the plate graphic will change from yellow to green, and the green Run Instrument arrow becomes enabled.

10. Click on the green Run Instrument arrow on the toolbar to start the sample run.

11. Monitor electrophoresis by observing the run, view, array or capillaries viewer windows in the data collection software. Each injection will take approximately 45 minutes.

5.B. Detection of Amplified Fragments Using the ABI PRISM® 3100 Genetic Analyzer with Data Collection Software, Version 1.0.1 or 1.1

Materials to Be Supplied by the User

- 95°C dry heating block, water bath or thermal cycler
- crushed ice or ice-water bath
- aerosol-resistant pipette tips
- 3100 capillary array, 36cm
- performance optimized polymer 4 (POP-4™) for the 3100
- 10X genetic analyzer buffer with EDTA
- MicroAmp® optical 96-well plate and septa for the 3100
- Hi-Di™ formamide (Applied Biosystems Cat.# 4311320)
- PowerPlex® Matrix Standards, 3100/3130 (Cat.# DG4650)

The quality of formamide is critical. Use Hi-Di™ formamide. Freeze formamide in aliquots at –20°C. Multiple freeze-thaw cycles or long-term storage at 4°C may cause breakdown of formamide. Poor-quality formamide may contain ions that compete with DNA during injection, which results in lower peak heights and reduced sensitivity. A longer injection time may not increase the signal.

Formamide is an irritant and a teratogen; avoid inhalation and contact with skin. Read the warning label, and take appropriate precautions when handling this substance. Always wear gloves and safety glasses when working with formamide.

Sample Preparation

1. Prepare a loading cocktail by combining and mixing Internal Lane Standard 600 and Hi-Di™ formamide as follows:

   [(0.5µl ILS 600) × (# injections)] + [(9.5µl Hi-Di™ formamide) × (# injections)]

   Note: The volume of internal lane standard used in the loading cocktail can be increased or decreased to adjust the intensity of the size standard peaks. The optimal peak height for the 100-base fragment of the internal lane standard is 500–1,000RFU. If peak heights are too low, we recommend altering the formamide/internal lane standard mix to contain 1.0µl of ILS 600 and 9.0µl of Hi-Di™ formamide. If peak heights are too high, we recommend altering the loading cocktail to contain 0.25µl of ILS 600 and 9.75µl of formamide.

2. Vortex for 10–15 seconds to mix.

3. Pipet 10µl of formamide/internal lane standard mix into each well.

5.B. Detection of Amplified Fragments Using the ABI PRISM® 3100 Genetic Analyzer with Data Collection Software, Version 1.0.1 or 1.1 (continued)

4. Add 1µl of amplified sample (or 1µl of PowerPlex® Y Allelic Ladder Mix). Cover wells with appropriate septa.

   Note: Instrument detection limits vary; therefore, injection time, injection voltage or the amount of product mixed with loading cocktail may need to be adjusted. Use the Module Editor in the Tools menu to modify injection time or voltage in the run module. If peak heights are higher than desired, samples can be diluted in Gold ST★R 1X Buffer before mixing with loading cocktail. The use of too much template DNA may result in uneven allele peak heights across loci. For best results, use less DNA template in the amplification reactions or reduce the number of cycles in the amplification program by 2–4 cycles to achieve the desired signal intensity.

5. Centrifuge plate briefly to remove air bubbles from the wells.

6. Denature samples at 95°C for 3 minutes, then immediately chill on crushed ice or in an ice-water bath for 3 minutes. Denature samples just prior to loading the instrument.

Instrument Preparation

Refer to the *ABI PRISM® 3100 Genetic Analyzer User's Manual* for instructions on cleaning the blocks, installing the capillary array, performing a spatial calibration and adding polymer to the reserve syringe.

1. Open the ABI PRISM® 3100 Data Collection Software.

2. Change the "GeneScan36_POP4DefaultModule" module run time to 2,000 seconds.

3. Change the injection voltage to 3kV.

4. Change the injection time to 11 seconds.

   Note: Instrument sensitivities can vary. Injection time and voltage may be adjusted in the Module Manager. A suggested range for the injection time is 3–22 seconds and for the injection voltage is 1–3kV.

5. Save the module with a new name (e.g., GeneScan36_POP4PowerPlexY_3kV_11secs_2000). Use this as the initial run module for all runs.

6. Open a new plate record. Name the plate, and select "GeneScan". Select the plate size (96-well). Select "Finish".

7. Complete the plate record spreadsheet for the wells you have loaded. Enter appropriate information into the Sample Name and Color Info columns. For allelic ladder samples, insert the word "ladder" into the Color Info column for the blue, yellow and green dye colors. This information must be entered to successfully analyze data with the PowerTyper™ Y Macro (Release 2.0).

8. In the BioLIMS Project column, select "3100_Project1" from the drop-down menu.
9. In the Dye Set column, select "Z" from the drop-down menu.
10. When using the ABI PRISM® 3100 Data Collection Software, Version 1.0.1 or 1.1, select "GeneScan36_POP4PowerPlexY_3kV_11secs_2000" from the drop-down menu in the Run Module 1 column.
11. To collect the data without autoanalyzing, select "No Selection" in the Analysis Module 1 column. Analysis parameters can be applied after data collection and during data analysis using the GeneScan® analysis software.
12. Select "OK". This new plate record will appear in the pending plate records table on the plate setup page of the collection software.
13. Place samples in the instrument, and close the instrument doors.
14. Locate the pending plate record that you just created, and click once on the name.
15. Once the pending plate record is highlighted, click on the plate graphic that corresponds to the plate on the autosampler that contains your amplified samples to link the plate to the plate record.
16. When the plate record is linked to the plate, the plate graphic will change from yellow to green, the plate record moves from the pending plate records table to the linked plate records table, and the Run Instrument button becomes enabled.
17. Select "Run Instrument" on the toolbar to start the sample run.
18. Monitor electrophoresis by observing the run, status, array and capillary views windows in the collection software. Each injection will take approximately 45 minutes.

5.C. Detection of Amplified Fragments Using the ABI PRISM® 310 Genetic Analyzer

Materials to Be Supplied by the User

- 95°C dry heating block, water bath or thermal cycler
- 310 capillaries, 47cm × 50µm
- performance optimized polymer 4 (POP-4™)
- 10X genetic analyzer buffer with EDTA
- sample tubes and septa
- aerosol-resistant pipette tips
- Hi-Di™ formamide (Applied Biosystems Cat.# 4311320)
- PowerPlex® Matrix Standards, 310 (Cat.# DG4640)
- crushed ice or ice-water bath

The quality of formamide is critical. Use Hi-Di™ formamide. Freeze formamide in aliquots at –20°C. Multiple freeze-thaw cycles or long-term storage at 4°C may cause breakdown of formamide. Poor-quality formamide may contain ions that compete with DNA during injection, which results in lower peak heights and reduced sensitivity. A longer injection time may not increase the signal.

Formamide is an irritant and a teratogen; avoid inhalation and contact with skin. Read the warning label, and take appropriate precautions when handling this substance. Always wear gloves and safety glasses when working with formamide.

Sample Preparation

1. Prepare a loading cocktail by combining Internal Lane Standard 600 (ILS 600) and Hi-Di™ formamide as follows:

   [(1.0µl ILS 600) × (# injections)] + [(24.0µl Hi-Di™ formamide) × (# injections)]

   Note: The volume of internal lane standard used in the loading cocktail can be increased or decreased to adjust the intensity of the size standard peaks. The optimal peak height for the 100-base fragment of the internal lane standard is 500–1,000RFU. If peak heights are too high, we recommend altering the loading cocktail to contain 0.5µl of ILS 600 and 24.5µl of Hi-Di™ formamide.

2. Vortex for 10–15 seconds to mix.

3. Combine 25.0µl of prepared loading cocktail and 1.0µl of amplified sample.

   Note: Instrument detection limits vary; therefore, injection time, injection voltage or the amount of product mixed with loading cocktail may need to be adjusted. If peak heights are higher than desired, samples can be diluted in Gold ST★R 1X Buffer before mixing with loading cocktail. The use of too much template DNA may result in uneven allele peak heights across loci. For best results, use less template DNA in the amplification reactions or reduce the number of cycles in the amplification program by 2–4 cycles (i.e., 10/18 or 10/20 cycling).

4. Combine 25.0µl of prepared loading cocktail and 1.0µl of PowerPlex® Y Allelic Ladder Mix.
5. Centrifuge tubes briefly to remove air bubbles from the wells.
6. Denature samples and ladder by heating at 95°C for 3 minutes, and immediately chill on crushed ice or in an ice-water bath for 3 minutes. Denature samples just prior to loading.
7. Place the tubes in the appropriate autosampler tray.
8. Place the autosampler tray in the instrument, and close the instrument doors.

Instrument Preparation

Refer to the instrument users' manual for instructions on cleaning the pump block, installing the capillary, calibrating the autosampler and adding polymer to the syringe.

1. Open the ABI PRISM® 310 Data Collection Software.
2. Prepare a GeneScan® sample sheet as described in the *ABI PRISM® 310 Genetic Analyzer User's Manual*. Enter the appropriate sample information in the Sample Info column.

   For rows containing PowerPlex® Y Allelic Ladder Mix, insert the word "ladder" in the Sample Info column for the blue dye color, yellow dye color and green dye color. This information must be entered to successfully analyze your data using the PowerTyper™ Y Macro.
3. Create a new GeneScan® injection list. Select the appropriate sample sheet by using the drop-down menu.

5.C. Detection of Amplified Fragments Using the ABI PRISM® 310 Genetic Analyzer (continued)

4. Select the "GS STR POP4 (1ml) A" Module using the drop-down menu. Change the injection time to the appropriate setting and the run time to 27 minutes. Keep the settings for the remaining parameters as shown below:

Inj. Secs:	2–5
Inj. kV:	15.0
Run kV:	15.0
Run °C:	60
Run Time:	27

   You may need to optimize the injection time for individual instruments. Injection times of 2–5 seconds are recommended for samples that contain 0.5–1ng of male template DNA. Allelic ladder and samples amplified with less than 32 cycles may work best with longer injection times (5 seconds). Use of highly sensitive instrumentation, amplification of >1ng male template, or use of 32 cycles may require shorter injection times.

   Note: Migration of fragments may vary slightly over the course of a long ABI PRISM® 310 Genetic Analyzer run. This may be due to changes in temperature or changes in the column. When analyzing many samples, injections of allelic ladder at different times throughout the run can aid in accurately genotyping the samples.

5. Select the appropriate matrix file (Section 3.B).

6. To analyze data automatically, select the auto analyze checkbox and the appropriate analysis parameters and size standard. Refer to the *ABI PRISM® 310 Genetic Analyzer User's Manual* for specific information on these options.

7. After loading the sample tray and closing the doors, select "Run" to start the capillary electrophoresis system.

8. Monitor electrophoresis by observing the raw data and status windows. Each sample will take approximately 35 minutes for syringe pumping, sample injection and sample electrophoresis.

5.D. Detection of Amplified Fragments Using the ABI PRISM® 377 DNA Sequencer

Materials to Be Supplied by the User

(Solution compositions are provided in Section 9.E.)

- Long Ranger® gel solution (Cambrex Cat.# 50611) or Long Ranger Singel® pack for ABI 377-36cm (Cambrex Cat.# 50691)
- 10% Ammonium Persulfate (Cat.# V3131)
- TEMED
- TBE 10X buffer
- Nalgene® tissue culture filter (0.2 micron)
- 36cm front and rear glass plates
- 36cm gel spacers (0.2mm thick)
- 36-well sharkstooth comb or 34-well square-tooth comb (0.2mm thick)
- clamps (e.g., large office binder clamps)
- gel-loading pipette tips
- aerosol-resistant pipette tips (Section 9.F)
- Liqui-Nox® or other detergent
- PowerPlex® Matrix Standards, 310 (Cat.# DG4640)
- Blue Dextran Loading Solution (Cat.# DV4351)
- crushed ice or ice-water bath
- 95°C dry heating block, water bath or thermal cycler

Acrylamide (Long Ranger® gel solution) is a neurotoxin and suspected carcinogen; avoid inhalation and contact with skin. Read the warning label, and take appropriate precautions when handling this substance. Always wear gloves and safety glasses when working with acrylamide solutions.

Polyacrylamide Gel Preparation

The following protocol is for preparation of a 36cm denaturing polyacrylamide gel for use with the ABI PRISM® 377 DNA Sequencer. Low-fluorescence glass plates are recommended and may be obtained from the instrument manufacturer.

1. Thoroughly clean glass plates with hot water and a 1% Liqui-Nox® solution. Rinse extremely well using deionized water. Allow the glass plates to air-dry in a dust-free environment.

2. Assemble glass plates by placing 0.2mm side gel spacers between the front and rear glass plates. Hold the plates together using binder clamps (4 clamps on each side). Place the assembly horizontally on a test tube rack or similar support.

5.D. Detection of Amplified Fragments Using the ABI PRISM® 377 DNA Sequencer (continued)

3. Prepare a 5% Long Ranger® acrylamide gel (total of 50ml) by combining the ingredients listed in Table 3. Stir the solution until the urea has dissolved.

Table 3. Preparation of a 5% Long Ranger® Polyacrylamide Gel.

Component	5% Gel	Final Concentration
urea	18g	6M
deionized water	26ml	–
10X TBE	5ml	1X
50% Long Ranger® gel solution	5ml	5%
total volume	50ml	

Note: Long Ranger Singel® Packs may be used.

4. Filter the acrylamide solution through a 0.2 micron filter (e.g., Nalgene® tissue culture filter), and degas for 5 minutes.

5. Add 35µl of TEMED and 250µl of fresh 10% ammonium persulfate to 50ml of acrylamide solution, and mix gently.

6. Using a disposable 60cc syringe, pour the gel by starting at the well end of the plates and carefully injecting the acrylamide between the horizontal glass plates. Allow the solution to fill the top width of the plates. While maintaining a constant flow of solution, gently tap the glass plates to assist movement of solution to the bottom of the plates and to prevent formation of bubbles.

7. Insert a 36-well sharkstooth comb or 34-well square-tooth comb between the glass plates. Sharkstooth combs with 64 or 96 wells also may be used.

8. Secure the comb with 3 evenly spaced clamps.

9. Keep the remaining acrylamide solution as a polymerization control.

10. Allow polymerization to proceed for at least 2 hours. Check the polymerization control to be sure that polymerization has occurred.

    Note: The gel may be stored overnight if a paper towel saturated with deionized water and plastic wrap are placed around the top and bottom to prevent the gel from drying out (crystallization of the urea will destroy the gel).

Instrument Preparation

1. Open the ABI PRISM® 377 Data Collection Software.
2. Prepare a sample sheet as described in the *GeneScan® Analysis Software User's Manual*. Enter the appropriate sample information in the Sample Info column.

   For lanes containing PowerPlex® Y Allelic Ladder Mix, insert the word "ladder" in the Sample Info column for the blue dye color, yellow dye color and green dye color. This information must be entered to successfully analyze your data using the PowerTyper™ Y Macro (Release 2.0).
3. Create a new GeneScan® run, and use the following settings:

Plate Check Module:	Plate Check A
PreRun Module:	PR GS 36A-2400
Run Module:	GS 36A-2400
Collect time:	2.5 hours
Well-to-Read distance:	36cm
4. Select the appropriate sample sheet and comb selection by using the drop-down menus.
5. Select the appropriate gel matrix file (Section 3.B).

Gel Pre-Run

1. Remove clamps from the polymerized acrylamide gel. If necessary, clean any excess acrylamide from the glass plates with paper towels saturated with deionized water.
2. Shave any excess polyacrylamide away from the comb, and remove the comb. If using a sharkstooth comb, carefully insert the sharkstooth comb teeth into the gel approximately 1–2mm.
3. Position the gel/glass plate unit in the 377 cassette.
4. Secure the cassette in the instrument, and perform a plate check as recommended in the *ABI PRISM® 377 DNA Sequencer User's Manual*. If the horizontal line graph is not flat, remove the cassette, clean the plate surface and repeat the plate check.
5. Add TBE 1X buffer to the top and bottom buffer chambers of the instrument.
6. Using a 60cc syringe filled with buffer, remove any air bubbles from the well area of the gel, and place the lid on the upper buffer chamber. Using a syringe fitted with a bent 18-gauge needle, remove any air bubbles from the bottom of the gel.
7. Attach the heating plate, connect the water tubing, attach all electrodes, close the instrument door and select "PreRun". Allow the gel to pre-run for 15–20 minutes or until the gel temperature is at least 40°C. Open the status window to monitor the gel temperature.
8. Prepare samples and allelic ladder samples during the gel pre-run.

5.D. Detection of Amplified Fragments Using the ABI PRISM® 377 DNA Sequencer (continued)

Sample Preparation and Loading

1. Prepare a loading cocktail by combining and mixing ILS 600 and Blue Dextran Loading Solution as follows:

   [(0.5µl ILS 600) × (# lanes)] + [(1.5µl Blue Dextran Loading Solution) × (# lanes)]

   Note: The volume of internal lane standard used in the loading cocktail can be increased or decreased to adjust the intensity of the size standard peaks.

2. Vortex for 10–15 seconds to mix.

3. Combine 2.0µl of prepared loading cocktail and 1.0µl of amplified sample.

   Note: Instrument detection limits vary; therefore, the amount of product mixed with loading cocktail may need to be adjusted. If peak heights are higher than desired, samples can be diluted in Gold ST★R 1X Buffer before mixing with loading cocktail. The use of too much template DNA may result in uneven allele peak heights across loci. For best results, use less template DNA in the amplification reactions or reduce the number of cycles in the amplification program by 2–4 cycles (i.e., 10/18 or 10/20 cycling).

4. Combine 2.0µl of prepared loading cocktail and 1.0µl of PowerPlex® Y Allelic Ladder Mix. Vortex the allelic ladder mix prior to pipetting.

5. Briefly centrifuge samples to bring the contents to the bottom of the tubes.

6. Just prior to loading the gel, denature samples by heating at 95°C for 3 minutes, and immediately chill on crushed ice or in an ice-water bath. Denature samples just prior to loading the gel.

7. After the 15- to 20-minute pre-run, pause the instrument by selecting "Pause". By pausing the pre-run, the water will continue to circulate, keeping the gel warm during sample loading.

8. Use a 60cc syringe filled with buffer and fitted with a bent 18-gauge needle to flush urea from the well area.

9. Load 1.5µl of each denatured sample into the respective wells.

   You may need to optimize the volume of sample loaded for individual instruments. We recommend loading volumes of 1.0–2.0µl.

10. Place the lid on the upper buffer chamber, and close the instrument door.

Gel Electrophoresis and Detection

1. After loading, select "Cancel" to stop the pre-run. Make sure that the run time is set at 2.5 hours, then select "Run" to begin electrophoresis.
2. Monitor electrophoresis by observing the gel image and status windows.
3. Allow electrophoresis to proceed for 2.5 hours.
4. Track and extract the gel lanes.

Reuse of Glass Plates

Separate the glass plates, and discard the gel. Clean glass plates with hot water and a detergent such as 1% Liqui-Nox® detergent. Rinse extremely well with deionized water, and allow the plates to air-dry. Do not scrape plates with abrasive materials during this process.

Note: Soap and oil may build up on plates, resulting in gel extrusion or hazy background. Soak plates in 2N HCl for 15 minutes, then rinse thoroughly to remove any buildup.

6. Data Analysis

6.A. PowerPlex® Panels and Bins Text Files with GeneMapper® *ID* Software, Version 3.2

To facilitate analysis of data generated with the PowerPlex® Y System, we have created panels and bins text files to allow automatic assignment of genotypes using GeneMapper® *ID* software, version 3.2. We recommend that users of GeneMapper® *ID* software, version 3.2, complete the *Applied Biosystems GeneMapper®* ID *Software Human Identification Analysis Tutorial* to familiarize themselves with proper operation of the software. For GeneMapper® *ID* software, version 3.1, users we recommend upgrading to version 3.2.

Getting Started

1. Obtain the proper panels and bins text files for use with GeneMapper® *ID* from the Promega web site at:
   www.promega.com/resources/tools/genemapper-id-software-panels-and-bin-sets/
2. Enter your contact information, and select "GeneMapper ID". Select "Submit".
3. Select the "PowerPlex® Panels & Bin Sets" link, and save the .zip file to your computer.
4. Open the files using the Windows® WinZip program, and save the unzipped files to a known location on your computer.

6.A. PowerPlex® Panels and Bins Text Files with GeneMapper® *ID* Software, Version 3.2 (continued)

Importing Panels and Bins Text Files

For detailed instructions, see the Applied Biosystems GeneMapper® *ID* software tutorial.

1. Open the GeneMapper® *ID* software, version 3.2.
2. Select "Tools", then "Panel Manager".
3. Highlight the Panel Manager icon in the upper left navigation pane.
4. Select "File", then "Import Panels".
5. Navigate to the saved panels and bins text files. Select "Promega_Panels_ID3.2.X.txt", where "X" refers to the most recent version of the panels and bins text files. Select "Import".
6. In the navigation pane, highlight the Promega_Panels_ID3.2.X folder that you just imported.
7. Select "File", then "Import Bin Set".
8. Navigate to the saved panels and bins text files. Select "Promega_Bins_ID3.2.X.txt", then "Import".
9. At the bottom of the Panel Manager window, select "OK". The Panel Manager window will close automatically.

6.B. Creating a Casework Analysis Method with GeneMapper® *ID* Software, Version 3.2

For detailed instructions, see the Applied Biosystems GeneMapper® *ID* software tutorial.

1. Select "Tools", then "GeneMapper Manager".
2. Select the Analysis Methods tab.
3. Select "New", and a new analysis method dialog box will open.
4. Select "HID", and select "OK".

   Note: If you do not see the HID option, you do not have the GeneMapper® *ID* software. Contact Applied Biosystems.
5. Enter a descriptive name for the analysis method, such as "PowerPlexY advanced".
6. Select the Allele tab (Figure 3).
7. Select the bins text file corresponding to the PowerPlex® System, "Promega_Bins_ID3.2.X", where "X" refers to the most recent version of the bin set.
8. Ensure that the "Use marker-specific stutter ratio if available" box is checked.

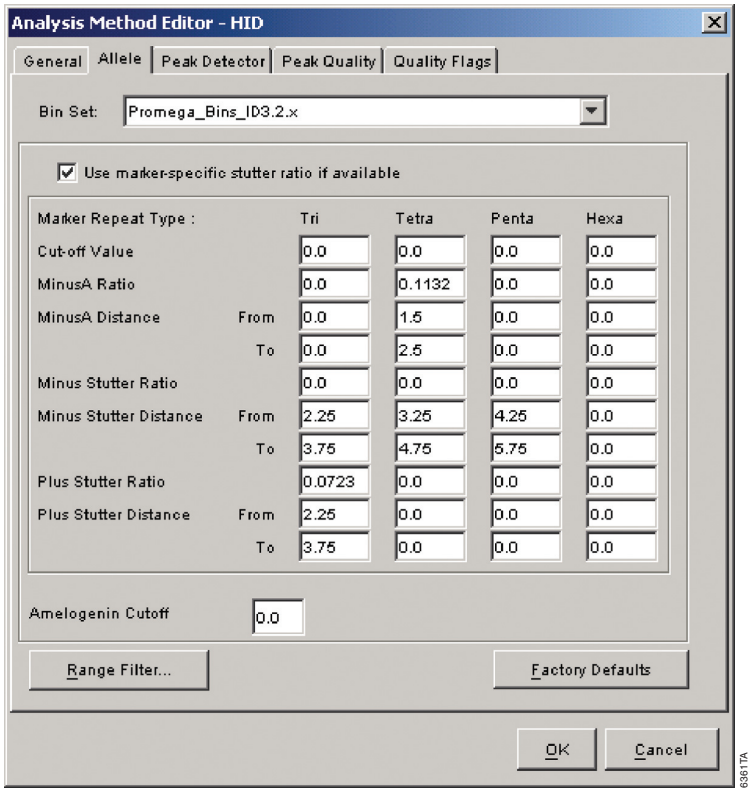

Figure 3. The Allele tab. Select the bins text file "Promega_Bins_ID3.2.X.txt", where "X" refers to the most recent version of the bins text file.

9. Enter the values shown in Figure 3 for proper filtering of stutter peaks when using the PowerPlex® Y System. For an explanation of the proper usage and effects of these settings, refer to the Applied Biosystems User Bulletin titled "*Installation Procedures and New Features for GeneMapper* ID *Software 3.2*".

   Note: Some of these settings have been optimized and are different from the recommended settings in the user bulletin.

10. Select the Peak Detector tab. We recommend the settings shown in Figure 4.

    Note: Select full range or partial range for the analysis range. When using a partial range, choose an appropriate analysis range based on your data. Choose a start point after the primer peak and just before the first defined internal lane standard peak to help ensure proper sizing of the internal lane standard.

11. Select the Peak Quality tab. You may change the settings for peak quality.

    Note: For Steps 11 and 12, see the GeneMapper® *ID* user's manual for more information.

12. Select the Quality Flags tab. You may change these settings.

13. Select "OK" to save your settings.

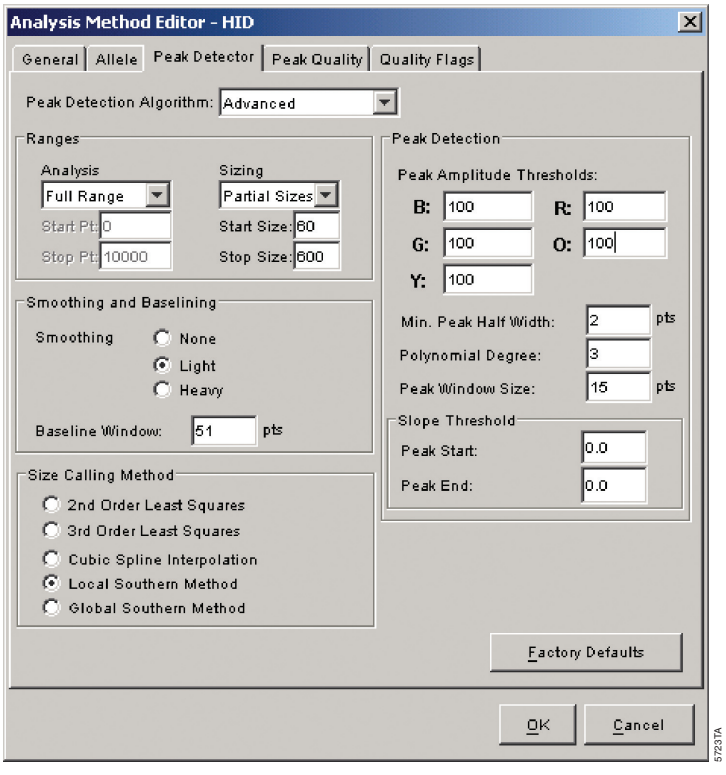

Figure 4. The Peak Detector tab.

Creating a Size Standard

1. Select "Tools", then "GeneMapper Manager".
2. Select the Size Standard tab.
3. Select "New".
4. Select "Basic or Advanced" (Figure 5). The type of analysis method selected must match the type of analysis method created earlier. Select "OK".
5. Enter a detailed name such as "ILS 600 advanced" in the Size Standard Editor (Figure 6).
6. Choose "Red" for the Size Standard Dye.
7. Enter the sizes of the internal lane standard fragments (see Section 9.C, Figure 13).
8. Select "OK".

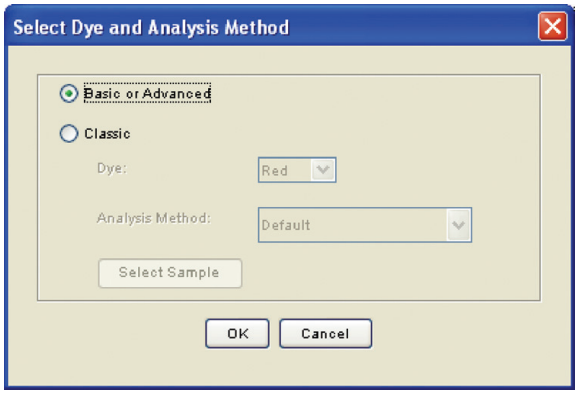

Figure 5. The Select Dye and Analysis Method window.

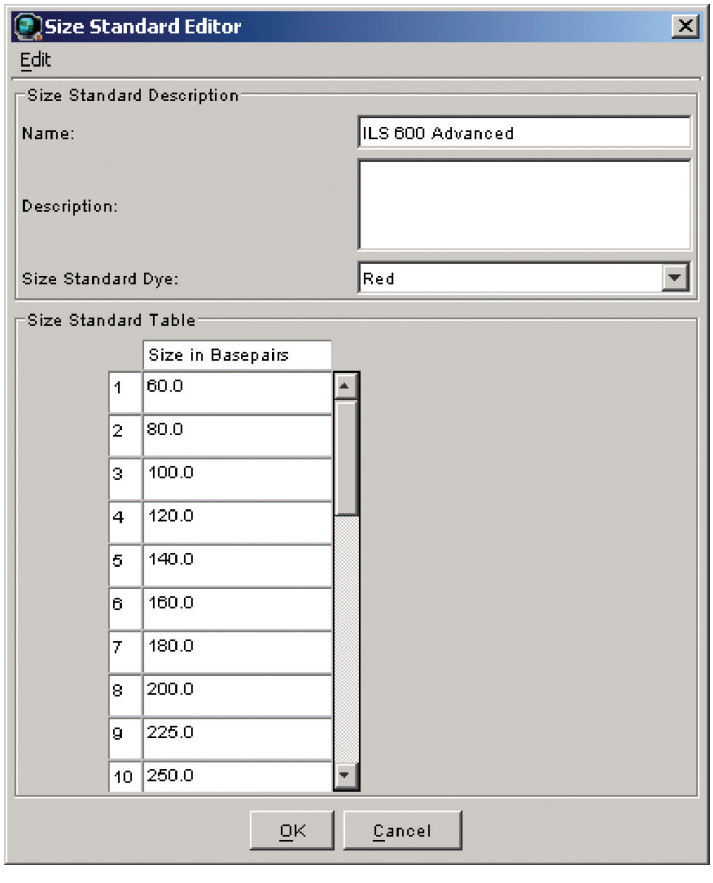

Figure 6. The Size Standard Editor.

6.B. Creating a Casework Analysis Method with GeneMapper® *ID* Software, Version 3.2 (continued)

Processing Sample Data for Casework

1. Import sample files into a new project as described in the *Applied Biosystems GeneMapper®* ID *Software Human Identification Analysis Tutorial.*
2. In the Sample Type column, use the drop-down menu to select "Ladder", "Sample", "Positive Control" or "Negative Control". Every folder in the project must contain at least one allelic ladder that is designated as such for proper genotyping.
3. In the Analysis Method column, select the analysis method created previously in the Creating a Casework Analysis Method section.
4. In the Panel column, select "PowerPlex_Y_ID3.2.X", where "X" refers to the most recent version of the panels files. This is the panels text file that was imported in Section 6.A.
5. In the Size Standard column, select the size standard that was created in Creating a Size Standard section.
6. If analyzing data from an ABI PRISM® 310 Genetic Analyzer or an ABI PRISM® 377 DNA Sequencer, ensure that the appropriate matrix file is selected in the Matrix column.
7. Select "Analyze" (green arrow button) to start data analysis.

6.C. Creating a Databasing or Paternity Analysis Method with GeneMapper® *ID* Software, Version 3.2

1. Select "Tools", then "GeneMapper Manager".
2. Select the Analysis Methods tab.
3. Select "New", and a new analysis method dialog box will open.
4. Select "HID", and select "OK".

   Note: If you do not see the HID option, you do not have the GeneMapper® *ID* software. Contact Promega Technical Services by e-mail: **genetic@promega.com** for assistance.
5. Enter a descriptive name for the analysis method, such as "PowerPlexY_20%filter".
6. Select the Allele tab.
7. Select the bins text file corresponding to the PowerPlex® System "Promega_Bins_ID3.2.X", where "X" refers to the most recent version of the bins text file.
8. Ensure that the "Use marker-specific stutter ratio if available" box is checked.

9. Enter the values shown in Figure 7 for proper filtering of peaks when using the PowerPlex® Y System. For an explanation of the proper usage and effect of these settings, refer to the Applied Biosystems user bulletin titled "*Installation Procedures and New Features for GeneMapper* ID *Software* 3.2".

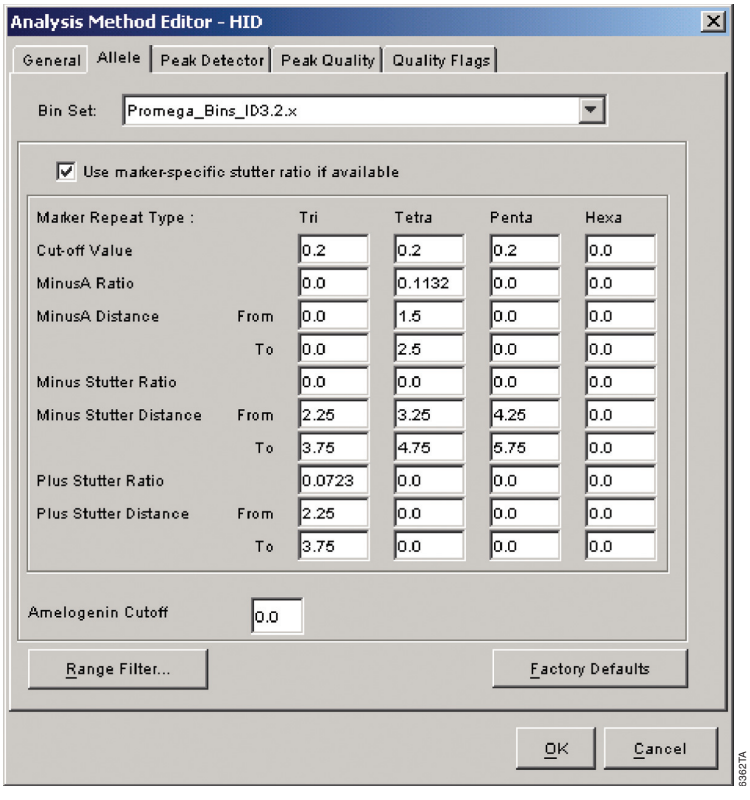

Figure 7. The Allele tab with settings for using a 20% peak filter. Select the bins text file "Promega_Bins_ID3.2.X.txt", where "X" refers to the most recent version of the bins text file.

Creating a Size Standard

1. Select "Tools", then "GeneMapper Manager".
2. Select the Size Standard tab.
3. Select "New".
4. Select "Basic or Advanced" (Figure 5). The type of analysis method selected must match the type of analysis method created earlier. Select "OK".
5. Enter a detailed name such as "ILS 600 advanced" in the Size Standard Editor (Figure 6).
6. Choose "Red" for the Size Standard Dye.
7. Enter the sizes of the internal lane standard fragments (see Section 9.C, Figure 13).
8. Select "OK".

6.C. Creating a Databasing or Paternity Analysis Method with GeneMapper® *ID* Software, Version 3.2 (continued)

Processing Sample Data for Databasing or Paternity Samples

1. Import sample files into a new project as described in the *Applied Biosystems GeneMapper®* ID *Software Human Identification Analysis Tutorial.*
2. In the Sample Type column, use the drop-down menu to select "Ladder", "Sample", "Positive Control" or "Negative Control". Every folder in the project must contain at least one ladder that is designated as such for proper genotyping.
3. In the Analysis Method column, select the analysis method created in the Creating a Databasing or Paternity Analysis Method section.
4. In the Panel column, select "PowerPlex_Y_ID3.2.X", where "X" refers to the most recent version of the panels files. This is the panels text file that was imported in Section 6.A.
5. In the Size Standard column, select the size standard that was created in the Creating a Size Standard section.
6. If analyzing data from an ABI PRISM® 310 Genetic Analyzer or an ABI PRISM® 377 DNA Sequencer, ensure that the appropriate matrix file is selected in the Matrix column.
7. Select "Analyze" (green arrow button) to start the data analysis.

6.D. Sample Analysis Using the GeneScan® Software and Windows® Operating Systems

1. Analyze data using the GeneScan® analysis software.
2. Review the raw data for one or more sample runs. Highlight the sample file name, then in the Sample menu, select "raw data". Move the cursor so the crosshair is on the baseline to the right of the large primer peak (before the first internal lane standard peak [red]). Use the X-value number shown at the bottom left of the window for the start position in the analysis parameters.
3. The recommended analysis parameters are shown in Figure 8.
4. The analysis parameters can be saved in the Params folder; in most installations this is located at:
   C:\AppliedBio\Shared\Analysis\Sizecaller\Params
5. Apply the stored analysis parameters file to the samples.
6. Assign a new size standard. Select a sample file, and highlight the arrow next to size standard. Select "define new". Assign the size standard peaks as shown in Figure 13 in Section 9.C. Store the size standard in the Size Standards folder at:
   C:\AppliedBio\Shared\Analysis\Sizecaller\SizeStandards
7. Apply the size standard file to the samples, then analyze the sample files. See Section 6.F for additional information on the use of the PowerTyper™ Y Macro (Release 2.0) and Genotyper® software.

Notes:

1. Peak heights outside the linear range of the instrument may generate artifact peaks due to instrument saturation (i.e., overloading the sample). Bleedthrough (pull-ups) from one color to another may be observed. Saturated signal may appear also as two peak (split peak).
2. If peak heights are not within the linear range of detection of the instrument, the ratio of stutter peaks to real allele peaks increases, and allele designations become difficult to interpret. The balance of peak heights also may appear less uniform.
3. There may be variation between instruments regarding the relative fluorescent units detected using the same sample. Furthermore, different instruments vary in the relative efficiency of color detection, affecting the dye color-to-dye color balance.

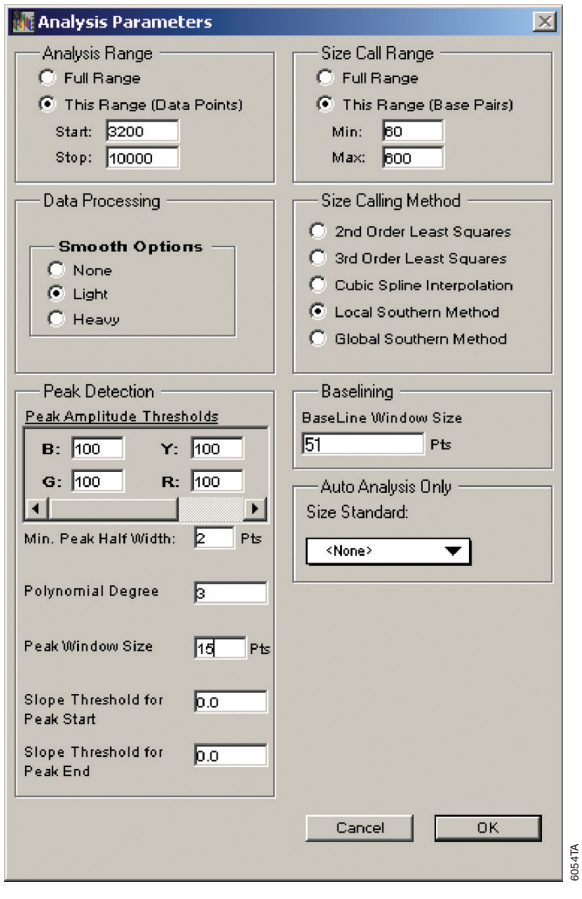

Figure 8. The Analysis Parameters window. The start point of the analysis range, which will vary, is defined in Section 6.D, Step 2.

6.E. Sample Analysis Using the GeneScan® Software and Macintosh® Operating Systems

1. Analyze data using the GeneScan® analysis software.
2. Review the raw data for one or more sample runs. Highlight the sample file name, then in the Sample menu select "raw data". Move the cursor so the crosshair is on the baseline to the right of the large primer peak (before the first internal lane standard peak [red]). Use the X-value number shown at the bottom left of the window for the start position in the analysis parameters.
3. The recommended analysis parameters are:

Analysis Range	Start: Defined in Step 2 Stop: 10,000
Data Processing	Baseline: Checked Multicomponent: Checked Smooth Options: Light[1]
Peak Detection	Peak Amplitude Thresholds[2]: B: Y: G: R: Min. Peak Half Width: 2pts
Size Call Range	Min: 60 Max: 600
Size Calling Method	Local Southern Method
Split Peak Correction	None

[1]Smooth options should be determined by individual laboratories.

[2]The peak amplitude thresholds are the minimum peak heights at which the software will call a peak. Values for the peak amplitude thresholds are usually 50–200RFU and should be determined by individual laboratories.

4. The analysis parameters can be saved in the Params folder.
5. Apply the stored analysis parameters file to the samples.
6. Assign a new size standard. Select a sample file, highlight the arrow next to size standard, then select "define new". Assign the size standard peaks as shown in Figure 13 in Section 9.C. Store the size standard in the Size Standards folder.
7. Apply the size standard file to the samples, then analyze the sample files. See Section 6.F for additional information on the use of the PowerTyper™ 16 Macro (Release 2.0) and Genotyper® software.

   For additional information regarding the GeneScan® analysis software, refer to the *GeneScan® Analysis Software User's Manual*.

Notes:

1. Peak heights outside the linear range of the instrument may generate artifact peaks due to instrument saturation (i.e., overloading the sample). Bleedthrough (pull-ups) from one color to another may be observed. Saturated signal may appear also as two peaks (split peak).
2. If peak heights are not within the linear range of detection of the instrument, the ratio of stutter peaks to real allele peaks increases, and allele designations become difficult to interpret. The balance of peak heights also may appear less uniform.
3. There may be variation between instruments regarding the relative fluorescent levels detected using the same sample. Furthermore, different instruments vary in the relative efficiency of color detection, affecting the dye color-to-dye color balance.

6.F. Sample Analysis Using the Genotyper® Software and PowerTyper™ Y Macro

To facilitate analysis of data generated with the PowerPlex® Y System, we have created a file to allow automatic assignment of genotypes using the Genotyper® software. After samples are amplified, detected using the ABI PRISM® 310 or 3100 Genetic Analyzer (using Data Collection Software, Version 1.0.1 or 1.1) or ABI PRISM® 377 DNA Sequencer and analyzed using the GeneScan® analysis software, sample files can be imported into the Genotyper® program and analyzed using the PowerTyper™ Y Macro (Release 2.0).

The PowerTyper™ Y Macro (Release 2.0) can be downloaded from the Promega web site at: **www.promega.com/resources/tools/powertyper-macros/**

The PowerTyper™ Y Macro (Release 2.0) is used in conjunction with Macintosh® Genotyper® software, version 2.5, and Windows NT® Genotyper® software, version 3.6, or later. The Genotyper® software must be installed on your computer before the PowerTyper™ Y Macro (Release 2.0) can be used.

Be certain the Sample Info (Macintosh® computers) or Color Info (Windows NT® operating systems) column for each lane containing allelic ladder mix contains the word "ladder". The macro uses the word "ladder" to identify the sample file(s) containing allelic ladder. Sample info can be added or modified after importing into the PowerTyper™ Macro. Highlight the sample, then select "show dye/lanes window" in the Views menu.

1. Transfer the PowerTyper™ Y Macro (Release 2.0) to a designated location on your computer hard drive.
2. Open the Genotyper® software, then the PowerTyper™ Y Macro. For questions about the Genotyper® software, refer to the *Genotyper® Analysis Software User's Manual*.

6.F. Sample Analysis Using the Genotyper® Software and PowerTyper™ Y Macro (continued)

3. In the File menu, select "Import", and import the GeneScan® project or sample files to be analyzed. Import the blue, yellow, green and red dye colors.

   Note: To select the dye colors to be imported, select "Set Preferences" in the Edit menu.

4. Double-click on the Check ILS macro. The macros are listed at the bottom left corner of the active window. A plots window will be displayed to show the internal lane standard (i.e., ILS 600) in the red dye color. Scroll down to view, and confirm that the internal lane standard fragment sizes are correct. If necessary, re-analyze samples using the GeneScan® software and redefine internal lane standard fragments.

   Note: The software uses one ladder sample to determine allele sizes. The macro uses the first ladder sample imported for allele designations.

5. For casework, double-click on the POWER macro. The POWER macro identifies alleles in the ladder sample and calculates offsets for all loci. This process may take several minutes. When completed, a plots window will open to display the allelic ladders (i.e., DYS391, DYS389I, DYS439, DYS389II, etc.).

   Alternatively, for databasing or paternity, double-click on the POWER 20% Filter macro. This macro has a higher level of filtering than the standard POWER macro to reduce the need for manual editing of peak labels. The POWER 20% Filter should not be used if mixtures may exist.

   In general, allelic ladders contain fragments of the same lengths as many known alleles for the locus. Allelic ladder sizes and repeat units are listed in Table 5 (Section 9.A). Analysis using GeneScan® analysis software and Genotyper® software allows allele determination by comparing amplified sample fragments with allelic ladders and internal lane standards. When using an internal lane standard, the calculated lengths of allelic ladder components may differ from those listed in the table. This is due to differences in migration resulting from sequence differences between the allelic ladder fragments and internal size standard and is not a matter of concern.

6. Double-click on the Allelic Ladders macro. A plots window will open to display the blue (fluorescein) dye allelic ladders (i.e., DYS391, DYS389I, DYS439 and DYS389II), green (JOE) dye allelic ladder (i.e., DYS438, DYS437, DYS19 and DYS392) and yellow (TMR) dye allelic ladders (i.e., DYS393, DYS390 and DYS385). Confirm that the correct allele designations were assigned to the allelic ladders (Figure 10 in Section 6.H).

   Note: The software uses one ladder sample to determine allele sizes. The macro uses the first ladder sample imported for allele designations. If the POWER macro is run a second time. the software will use the second ladder; if the POWER macro is run a third time, the software will use the third ladder, etc., until all ladders in the project are used. If an allelic ladder fails to be analyzed or if many off-ladder alleles are found in the samples, samples should be re-analyzed using another ladder from the project.

7. Double-click on the Display Fluorescein Data macro to display the blue dye for all sample injections or lanes. Scroll down to observe and edit as needed.

8. Double-click on the Display TMR Data macro to display the yellow dye for all sample injections or lanes. Scroll down to observe and edit as needed.

9. Double-click on the Display JOE Data macro to display the green dye for all sample injections or lanes. Scroll down to observe and edit as needed.

10. Create the appropriate table by selecting the PowerTable, Make Allele Table or Make Vertical Table macro. The three available table formats are shown below. The PowerTable option allows up to four alleles per sample file. Additional information such as low peak signal or high peak signal is also included. The Allele Table and Vertical Table options include only two alleles per locus. If more than two alleles are present at a locus, the smallest alleles identified are included. The Allele Table format displays the categories (loci) in columns, while the Vertical table format displays the categories in rows. These tables can be customized to fit needs. To save data in tables, go to the Table drop-down menu, highlight "Export to File..." and save the file with the desired name and location. The saved file can be viewed and analyzed using Microsoft® Excel.

PowerTable Format

Sample Info	Sample Comment	Category	Peak 1	Peak 2	Peak 3	Peak 4	Over-flow	Low Signal	Satura-tion	Edited Label	Edited Row

Allele Table Format

Sample Info	Category Allele 1	Category Allele 2	Category Allele 1	Category Allele 2	Category Allele 1	Category Allele 2	Category Allele 1	Category Allele 2

Vertical Table Format

Sample Info	Category	Peak 1	Peak 2

11. Save the analyzed data. Go to the File menu, and select "Save as".

The PowerTyper™ Macro is a Genotyper® file and can be overwritten if "Save" is used instead of "Save as".

6.G. Controls

1. Observe the results for the negative control. Using the protocols defined in this manual, the negative control should be devoid of amplification products.
2. Observe the results for the 2800M Control DNA. Compare the 2800M Control DNA allelic repeat sizes with the locus-specific allelic ladder. The expected 2800M Control DNA allele designations for each locus are listed in Table 5 (Section 9.A).

6.H. Results

Representative results of the PowerPlex® Y System are shown in Figure 9. The PowerPlex® Y Allelic Ladder Mix is shown in Figure 10.

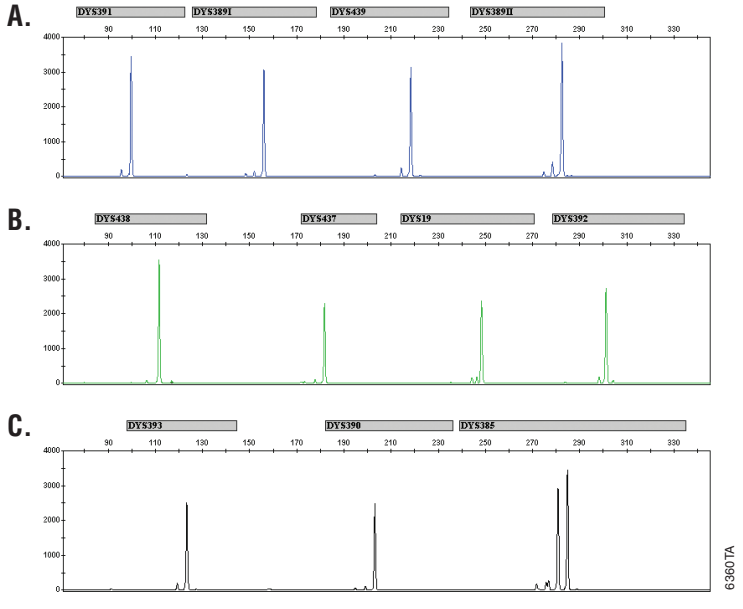

Figure 9. The PowerPlex® Y System. A single-source male template DNA (0.5ng) was amplified using the PowerPlex® Y System 10X Primer Pair Mix. Amplification products were detected using an Applied Biosystems 3130 Genetic Analyzer using a 3kV, 3-second injection. Results were analyzed using GeneMapper® *ID* software, version 3.2. **Panel A.** An electropherogram showing the peaks of the fluorescein-labeled loci DYS391, DYS389I, DYS439 and DYS389II. **Panel B.** An electropherogram showing the peaks of the JOE-labeled loci DYS438, DYS437, DYS19 and DYS392. **Panel C.** An electropherogram showing the peaks of the TMR-labeled loci DYS393, DYS390 and DYS385.

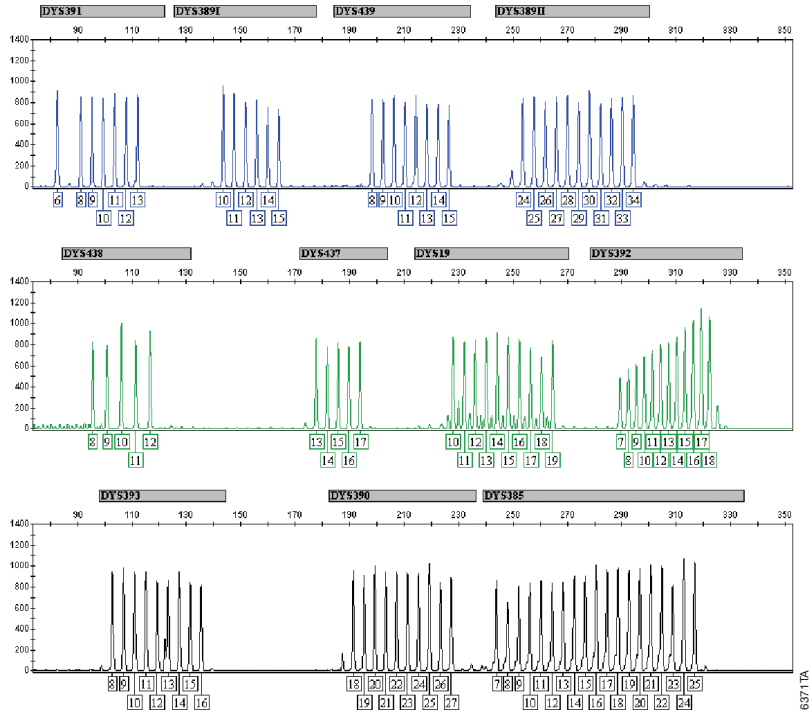

Figure 10. The PowerPlex® Y Allelic Ladder Mix. The PowerPlex® Y Allelic Ladder Mix was analyzed with an Applied Biosystems 3130 Genetic Analyzer using a 3kV, 3-second injection. The sample file was analyzed with GeneMapper® *ID* software, version 3.2, and PowerPlex® Y panels and bins text files. **Panel A.** The fluorescein-labeled allelic ladder components and their allele designations. **Panel B.** The JOE-labeled allelic ladder components and their allele designations. **Panel C.** The TMR-labeled allelic ladder components and their allele designations.

Stutter and Artifacts

Stutter products are a common amplification artifact associated with STR analysis. The pattern and intensity of stutter may differ between primer sets for the same loci due to reaction and amplification conditions and the labeled strand direction. For samples with increased signal or template amount, stutter products, which are one and occasionally two repeat units (Table 4) smaller than the true allele peak, are often observed. In addition to stutter peaks, several other stutter-like peaks can be observed at some PowerPlex® Y loci. The DYS392 locus and occasionally other loci with high signal or template amount may show a peak one repeat unit larger than the true allele. DYS19 and DYS389II can display low-level products in the n–2 and n+2 positions (two bases below and above the true allele peak, respectively), with the DYS19 n–2 product being the most prominent. DYS437 and DYS385 also may show low-level peaks in the n–5 position, with DYS437, DYS385 and DYS393 also displaying an n–9 to n–10 product.

The intensity of stutter and stutter-like peaks is directly related to signal intensity. Results may vary based on laboratory optimization. Internal laboratory validation should be performed.

A low-level artifact in the DYS438 region of the JOE channel may be observed between 114–120bp. This artifact is not template-derived and may appear in the negative control and in low-product-yield analyses. The peak height of this artifact may increase with longer injection time or higher injection voltage. In addition, low-level artifacts in the noncalling region between the DYS393 and DYS390 assay ranges of the TMR channel may be observed at approximately 150–165bp. These artifacts also are not template-derived and may appear in the negative control and in low-product-yield analyses. In amplified samples, noise below the allele calling (assay) range in the blue (<70bp), green (<90bp) and yellow (<95bp) channels can be observed. In the allelic ladder, low-level artifact peaks can be observed; most notable is a series of peaks above DYS389II (Figure 10). In general, none of these artifacts should affect interpretation due to the peak intensity and position relative to the allelic ladder peaks. However, their intensity can be decreased by reducing the injection time or loading volume, or signal thresholds can be increased during analysis to exceed the observed noise level.

DYS385a/b Concordance

Documentation of nonconcordance has been previously described for the DYS385a/b locus. An initial publication of primer sequences by Kayser (14) incorporates a single-base deletion between the primer binding sites but outside of the repeat region (11,13,21). An alternative set of primer sequences, published later by Schneider (22), produces a much smaller amplicon and is internal to this mutation site. Amplification of a sample with this rare single-base deletion in the DYS385a/b flanking region using the Kayser primers will consistently produce an amplicon that types one base shorter in length than that generated with the Schneider sequences. The Schneider primer sequences are commonly used (13).

The PowerPlex® Y System uses the Schneider primer-binding sites to allow high male specificity with a much smaller product size compared to the Kayser sites. Concordance/proficiency testing can be accomplished with the U.S. National Institute of Standards and Technology (NIST) Standard Reference Material® (SRM) 2395 Human Y-Chromosome DNA Profiling Standard (Gaithersburg, MD).

7. Troubleshooting

For questions not addressed here, please contact your local Promega Branch Office or Distributor. Contact information available at: **www.promega.com**. E-mail: **genetic@promega.com**

7.A. Amplification and Fragment Detection

Symptoms	Causes and Comments
Faint or absent allele peaks	Impure template DNA. Because of the small amount of template used, this is rarely a problem. Depending on the DNA extraction procedure used and sample source, inhibitors may be present in the DNA sample.
	Insufficient template. Use the recommended amount of male template DNA.
	Insufficient template. Low-copy-number (LCN) analysis using capillary electrophoresis may benefit from reducing competing charged particles during injection. This can be accomplished with post-PCR cleanup or desalting, lower-conductivity formamide or reduced amounts of ILS 600. Perform in-house validation for any of these methods.
	Insufficient enzyme activity. Use the recommended amount of AmpliTaq Gold® DNA polymerase. Check the expiration date on the tube label.
	Incorrect amplification program. Confirm the program.
	High salt concentration or altered pH. If the DNA template is stored in TE buffer that is not pH 8.0 or contains a higher EDTA concentration, the DNA volume should not exceed 20% of the total reaction volume. Carryover of K^+, Na^+, Mg^{2+} or EDTA from the DNA sample can negatively affect PCR. A change in pH also may affect PCR. Store DNA in TE^{-4} buffer (10mM Tris-HCl [pH 8.0], 0.1mM EDTA) or nuclease-free water.
	Thermal cycler, plate or tube problems. Review the thermal cycling protocols in Section 4.B. We have not tested other reaction tubes, plates or thermal cyclers. Calibrate the thermal cycler heating block, if necessary.
	Primer concentration was too low. Use the recommended primer concentration. Vortex the PowerPlex® Y 10X Primer Pair for 15 seconds to mix before use.
	Heat-denature samples for the recommended time, and cool on crushed ice or in an ice-water bath immediately prior to capillary electrophoresis.
	Poor capillary electrophoresis injection (ILS 600 peaks also affected). Re-inject the sample. Check the syringe for leakage. Check the laser power.
	Poor-quality formamide was used. Use only Hi-Di™ formamide when analyzing samples.

7.A. Amplification and Fragment Detection (continued)

Symptoms	Causes and Comments
Extra peaks visible in one or all color channels	Contamination with another template DNA or previously amplified DNA. Cross-contamination can be a problem. Use aerosol-resistant pipette tips, and change gloves regularly.
	Samples were not completely denatured. Heat denature samples for the recommended time, and cool on crushed ice or in an ice-water bath immediately prior to capillary electrophoresis. Do not cool the samples in a thermal cycler set at 4°C, as this may lead to artifacts due to DNA re-annealing.
	Artifacts of STR amplification. Amplification of STRs sometimes generates artifacts that appear as faint peaks one repeat unit smaller than the allele. Stutter band peak heights can be high if the samples are overloaded. See Section 6.H for additional information on stutter and artifacts.
	High background. Load less amplification product, or decrease injection time. See Section 5.
	Capillary electrophoresis (CE)-related artifacts ("spikes"). Minor voltage changes or urea crystals passing by the laser can cause "spikes" or unexpected peaks. Spikes sometimes appear in one color but often are easily identified by their presence in more than one color. Re-inject samples to confirm.
	CE-related artifacts (contaminants). Contaminants in the water used with the instrument or to dilute the 10X genetic analyzer buffer may generate peaks in the blue and green dye colors. Use autoclaved water; change vials and wash buffer reservoir.
	Excessive amount of DNA. Amplification of >2ng male DNA template can result in a higher number of stutter bands. Use less template DNA, or reduce the number of cycles in the amplification program by 2–4 cycles (10/20 or 10/18 cycling).
	Pull-up or bleedthrough. Pull-up can occur when peak heights are too high or if a poor or incorrect matrix has been applied to the samples. • For the ABI PRISM® 310 Genetic Analyzer and 377 DNA Sequencer, generate a new matrix, and apply it to the samples. For the ABI PRISM® 3100 and 3100-*Avant* Genetic Analyzers and Applied Biosystems 3130 and 3130*xl* Genetic Analyzers, perform a new spectral calibration and re-run the samples. • Instrument sensitivities can vary. Optimize the injection or gel loading conditions. See Section 5.
	Long-term storage of amplified sample in formamide can result in degradation. Repeat sample preparation using fresh formamide.
	The CE polymer was beyond its expiration date, or polymer was stored at room temperature for more than one week.
	Maintain instrumentation on a daily or weekly basis, as recommended by the manufacturer.

Symptoms	Causes and Comments
Allelic ladder not running the same as samples	Allelic ladder and primer pair mix were not compatible. Ensure that the allelic ladder is from the same system as the primer pair mix.
	Buffer incompatibility. Samples were diluted in the wrong buffer. Use Gold ST★R 1X Buffer to dilute samples.
	Poor-quality formamide. Use only Hi-Di™ formamide when analyzing samples.
	Be sure the allelic ladder and samples are from the same instrument run.
	Migration of samples changed slightly over the course of a CE run with many samples. This may be due to changes in temperature or the CE column over time. Use a different injection of allelic ladder to determine sizes.
	Poor injection of allelic ladder. Include more than one ladder per instrument run.
Peak height imbalance	Excessive amount of DNA. Amplification of >1ng of male template can result in an imbalance, with smaller loci showing more product than larger loci. Use less template, or reduce the number of cycles in the amplification program by 2–4 cycles (10/20 or 10/18 cycling) to improve locus-to-locus balance. **Note:** Dilution of overamplified samples can result in dropout of larger loci.
	Degraded DNA sample. DNA template was degraded, and larger loci show diminished yield. Repurify the template DNA.
	Insufficient male template DNA. Use the recommended amount of male template DNA. Stochastic effects can occur when amplifying low amounts of template.
	Miscellaneous balance problems. Thaw the 10X Primer Pair Mix and Gold ST★R 10X Buffer completely, and vortex for 15 seconds before using. Do not centrifuge the 10X Primer Pair Mix after mixing. Calibrate thermal cyclers and pipettes routinely. Using a 59°C annealing temperature instead of 60°C has been shown to improve balance in some instances.
	Impure template DNA. Inhibitors that may be present in forensic samples can lead to allele dropout or imbalance.
	Presence of female DNA. Female DNA at a concentration ≥100X that of the male component can decrease the relative yield of some loci.

7.B. GeneMapper® *ID* Analysis Software

Symptoms	Causes and Comments
Alleles not called	To analyze samples with GeneMapper® *ID* software, the analysis parameters and size standard must both have "Basic or Advanced" as the analysis type. If they are different, an error is obtained (Figure 11).
	An insufficient number of ILS 600 fragments was defined. Be sure to define at least one ILS 600 fragment smaller than the smallest sample peak or allelic ladder peak and at least one ILS 600 fragment larger than the largest sample peak or allelic ladder peak.
	Run was too short, and larger peaks in ILS were not captured. Not all ILS 600 peaks defined in the size standard were detected during the run. • Create a new size standard using the internal lane standard fragments present in the sample. • Re-run samples using a longer run time.
Off-ladder alleles	An allelic ladder from a different run than the samples was used. Re-analyze samples with an allelic ladder from the same run.
	The GeneMapper® *ID* software requires that the allelic ladder be imported from the same folder as the sample. Be sure that the allelic ladder is in the same folder as the sample. Create a new project and re-analyze, as described in Section 6.B or 6.C.
	Panels text file selected for analysis was incorrect for the STR system used. Assign correct panels file that corresponds to the STR system used for amplification.
	The allelic ladder was not identified as an allelic ladder in the Sample Type column.
	The wrong analysis type was chosen for the analysis method. Be sure to use the HID analysis type.
	The internal lane standard was not properly identified in the sample. Manually redefine the sizes of the size standard fragments in the sample.

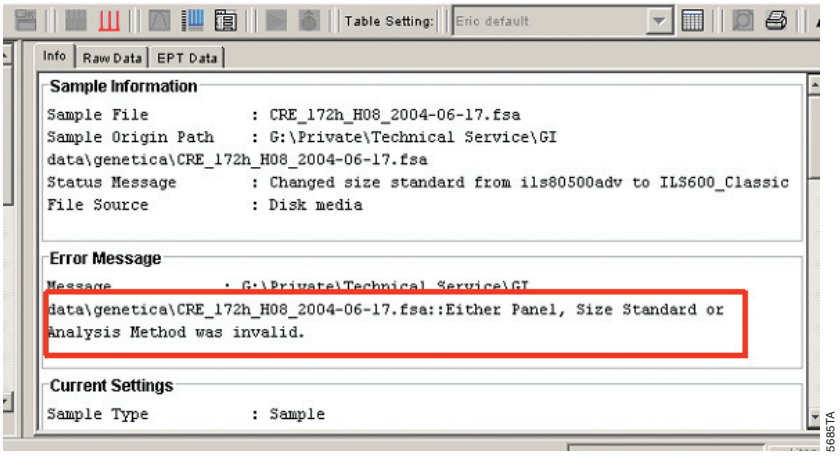

Figure 11. The error message that appears in the GeneMapper® *ID* software when the analysis parameters and size standard have different analysis types.

Symptoms	Causes and Comments
Size standard not called correctly (Figure 12)	Starting data point was incorrect for the partial range chosen in Section 6.B. Adjust the starting data point in the analysis method. Alternatively, use a full range for the analysis.
	Extra peaks in advanced mode size standard. Open the Size Match Editor. Highlight the extra peak, select "Edit" and select "delete size label". Select "auto adjust sizes".
	Run was too short, and larger peaks in ILS were not captured. Not all ILS 600 peaks defined in the size standard were detected during the run. • Create a new size standard using the internal lane standard fragments present in the sample. • Re-run samples using a longer run time.
Peaks in size standard missing	If peaks are below threshold, decrease the peak amplitude threshold in the analysis method for the red channel to include peaks.
	If peaks are low-quality, redefine the size standard for the sample to skip these peaks.
Error message: "Either panel, size standard, or analysis method is invalid"	The size standard and analysis method were not in the same mode ("Classic" vs. "Basic or Advanced"). Be sure both files are set to the same mode, either Classic or Basic or Advanced.
No alleles called, but no error message appears	Panels text file was not selected for sample. In the Panel column, select the appropriate panels text file for the STR system that was used.
	No size standard was selected. In the Size Standards column, be sure to select the appropriate size standard.
	Size standard was not correctly defined, or size peaks were missing. Redefine size standard to include only peaks present in your sample. Terminating analysis early or using short run times will cause larger ladder peaks to be missing. This will cause your sizing quality to be flagged as "red", and no allele sizes will be called.

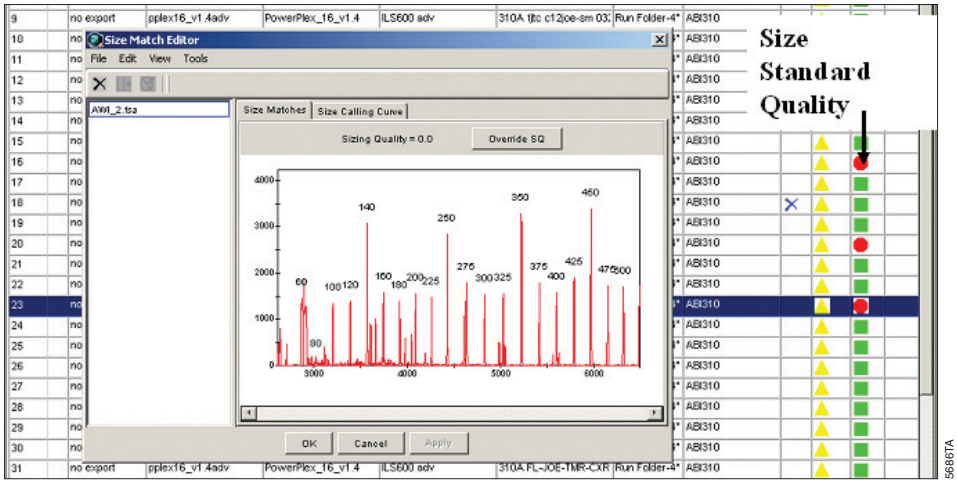

Figure 12. An example showing improper assignment of size standard fragments in the GeneMapper® *ID* software.

7.B. GeneMapper® *ID* Analysis Software (continued)

Symptoms	Causes and Comments
Error message: "Both the Bin Set used in the Analysis Method and the Panel must belong to the same Chemistry Kit".	The bins text file assigned to the analysis method may have been deleted. In the GeneMapper® Manager, select the Analysis Methods tab, and open the analysis method of interest. Select the Alleles tab, and select an appropriate bins text file.
	The wrong bins text file was chosen in the analysis method Allele tab. Be sure to choose the appropriate bins text file, as shown in Figure 3.
Significantly raised baseline	• Poor spectral calibration for the ABI PRISM® 3100 and 3100-*Avant* Genetic Analyzers and Applied Biosystems 3130 and 3130*xl* Genetic Analyzers. Perform a new spectral calibration and re-run the samples. • Poor matrix for the ABI PRISM® 310 Genetic Analyzer. Re-run and optimize the matrix.
	Use of Classic mode analysis method. Use of Classic mode analysis on samples can result in baselines with more noise than those analyzed using the Basic or Advanced mode analysis method. Advanced mode analysis methods and size standards are recommended.
Red bar appears during analysis of samples, and the following error message appears when data are displayed: "Some selected sample(s) do not contain analysis data. Those sample(s) will not be shown".	If none of the samples had matrices applied when run on the ABI PRISM® 310 Genetic Analyzer, no data will be displayed. Apply a matrix file during analysis in the GeneMapper® *ID* software and re-analyze.
Error message after attempting to import panels and bins text files: "Unable to save panel data: java.SQLEException: ORA-00001: unique constraint (IFA.CKP_NNN) violated".	There was a conflict between different sets of panels and bins text files. Check to be sure that the bins are installed properly. If not, delete all panels and bins text files, and re-import files in a different order.
Allelic ladder peaks are labeled off-ladder (OL)	GeneMapper® *ID* software was not used, or microsatellite analysis settings were used instead of HID analysis settings. GeneMapper® software does not use the same algorithms as GeneMapper® *ID* software and cannot correct for sizing differences using the allelic ladder. Promega recommends using GeneMapper® *ID* software for analysis of PowerPlex® reactions. If using GeneMapper® *ID* software, version 3.2, be sure that the analysis method selected is an HID method. This can be verified by opening the analysis method using the GeneMapper® Manager, then selecting the General tab. The analysis type cannot be changed. If the method is not HID, it should be deleted and a new analysis method created.

7.C. PowerTyper™ Y Macro

Symptoms	Causes and Comments
File does not open on your computer	Genotyper® software was not installed. Be certain that the Genotyper® software, version 2.5 (Macintosh®) or version 3.6 or higher (Windows NT®), is installed.
	Incorrect version of Genotyper® software. The PowerTyper™ Y Macro will not work with Genotyper® software versions prior to version 2.5.
Error message: "Could not complete the "Run Macro" command because no dye/lanes are selected"	Allelic ladder sample files were not identified. Be certain the Sample Info or Color Info column for each lane containing PowerPlex® Y Allelic Ladder Mix contains the word "ladder". The macro uses the word "ladder" to identify the sample files containing allelic ladder.
	All four dye colors were not imported. For Genotyper® software, versions 2.5 and 3.5 or higher, set preferences (in the Edit menu) to import the blue, green, yellow and red colors.
Error message: "Could not complete the "Run Macro" command because the labeled peak could not be found"	Peak heights for one or more alleles in the allelic ladder sample file were below 150RFU. The allelic ladder categories are defined as having a minimum peak height of 150RFU. If peak heights of ladder alleles are below 150RFU, the software will not be able to locate the allele peak. Re-run the allelic ladder using more sample or longer injection time to assure peak heights above 150RFU.
	CE spikes in the allelic ladder sample were identified as alleles by the macro. Use a different injection of allelic ladder.
	Allelic ladder data were not compatible with the PowerTyper™ file used. Confirm that the PowerTyper™ Macro file matches the allelic ladder being used.
	The base-pair size of alleles in the allelic ladder were outside of the defined category range. Be sure internal lane standard fragments are correctly sized. Redefine internal lane standard fragments, and re-analyze the sample using GeneScan® software. Compare the size of the smallest allele in the allelic ladder with the base-pair size and range listed in the categories for the same alleles. If necessary, increase the category start range (in the category window) to greater than ±6bp, and save the macro under a new name.
	Allelic ladder peaks were too high, causing stutter peaks to be called as allele peaks. Use a shorter injection time, decrease the amount of allelic ladder used or re-analyze the allelic ladder sample using increased peak amplitude thresholds in the GeneScan® analysis parameters.

Symptoms	Causes and Comments
The plots window or allele table does not display all data	The macros were not run in the proper order. Use the POWER or POWER 20% Filter macro option.
	All four dye colors were not imported. For Genotyper® software, versions 2.5 and 3.5 or higher, set preferences (in the Edit menu) to import the blue, green, yellow and red colors.
The Check ILS macro displays an empty plot window	All four dye colors were not imported. For Genotyper® software, versions 2.5 and 3.5 or higher, set preferences (in the Edit menu) to import the blue, green, yellow and red colors.
Off-ladder peaks	Migration of samples changed slightly over the course of a CE run with many samples. This may be due to changes in temperature or the CE column over time. Use a different injection of allelic ladder to determine sizes in the PowerTyper™ Y Macro (Release 2.0). Do not use the first injection on a new column for the ladder sample.
	The base-pair size of alleles was incorrect because incorrect fragment sizes were assigned to the internal lane standard. Confirm that internal lane standard fragments are assigned correctly. Re-analyze the sample using GeneScan® software, and redefine the internal lane standard fragments.

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Additional STR references can be found at: **www.promega.com/geneticidentity/**

9. Appendix

9.A. Advantages of Using the Loci in the PowerPlex® Y System

The loci included in the PowerPlex® Y System (Tables 4 and 5) were selected because they represent well characterized loci generally accepted for forensic use. This multiplex includes all loci in the "European minimal haplotype" (DYS19, DYS385a/b, DYS389I/II, DYS390, DYS391, DYS392 and DYS393) and the Scientific Working Group—DNA Analysis Methods (SWGDAM)-recommended Y-STR panel (European minimal haplotype plus DYS438 and DYS439) plus DYS437. More information on the European minimal haplotype can be found at: **www.ystr.org** (12).

The PowerPlex® Y System includes an extensive allelic ladder containing the most common variants observed at each locus. Table 5 lists the alleles in the allelic ladder and the haplotype of the 2800M Control DNA.

Table 4. The PowerPlex® Y System Locus-Specific Information.

STR Locus	Label	Chromosomal Location	GenBank® Accession Number	Repeat Sequence[1] 5´→3´
DYS391	FL	Yq	G09613	TCTA (14)
DYS389I/II	FL	Yq	AF140635	[TCTG][TCTA] Complex (14)
DYS439	FL	Yq	AC002992	GATA (23)
DYS393	TMR	Yq	G09601	AGAT (14)
DYS390	TMR	Yq	AC011289	[TCTG][TCTA] Complex (14)
DYS385a/b	TMR	Yq	Z93950	GAAA (14)
DYS438	JOE	Yq	AC002531	TTTTC (23)
DYS437	JOE	Yq	AC002992	[TCTA][TCTG] Complex (23)
DYS19	JOE	Yq	X77751	TAGA Complex (14)
DYS392	JOE	Yq	G09867	TAT (14)

[1]The August 1997 report (24,25) of the DNA Commission of the International Society for Forensic Haemogenetics (ISFH) states, "1) for STR loci within coding genes, the coding strand shall be used and the repeat sequence motif defined using the first possible 5´ nucleotide of a repeat motif; and 2) for STR loci not associated with a coding gene, the first database entry or original literature description shall be used".

TMR = carboxy-tetramethylrhodamine

FL = fluorescein

JOE = 6-carboxy-4´,5´-dichloro-2´,7´-dimethoxyfluorescein

Table 5. The PowerPlex® Y System Allelic Ladder Information.

STR Locus	Label	Size Range of Allelic Ladder Components[1] (bases)	Repeat Numbers of Allelic Ladder Components	Alleles Observed in 2800M Control DNA
DYS391	FL	90–118	6, 8–13	10
DYS389I	FL	148–168	10–15	14
DYS439	FL	203–231	8–15[2]	12
DYS389II	FL	256–296	24–34	31
DYS393	TMR	104–136	8–16	13
DYS390	TMR	191–227	18–27	24
DYS385	TMR	243–315	7–25	13,16
DYS438	JOE	101–121	8–12	9
DYS437	JOE	183–199	13–17	14
DYS19	JOE	232–268	10–19	14
DYS392	JOE	294–327	7–18	13

[1]When using an internal lane standard, such as the Internal Lane Standard 600, the calculated sizes of allelic ladder components may differ from those listed. This occurs because different sequences in allelic ladder and ILS components may cause differences in migration. The dye label also affects migration of alleles.

[2]Follows the original nomenclature described by Ayub *et al.* (23).

We have carefully selected primers to avoid or minimize artifacts, including those associated with *Taq* DNA polymerase, such as repeat slippage and terminal nucleotide addition. Repeat slippage (26,27), sometimes called "n–4 bands", "stutter" or "shadow bands", is due to the loss of a repeat unit during DNA amplification, somatic variation within the DNA, or both. The amount of this artifact observed depends primarily on the locus and the DNA sequence being replicated.

Terminal nucleotide addition (28,29) occurs when *Taq* DNA polymerase adds a nucleotide, generally adenine, to the 3´ ends of amplified DNA fragments in a template-independent manner. The efficiency with which this occurs varies with different primer sequences. Thus, an artifact band one base shorter than expected (i.e., missing the terminal addition) is sometimes seen. We have modified primer sequences and added a final extension step of 60°C for 30 minutes (30) to the amplification protocol to provide conditions for essentially complete terminal nucleotide addition when recommended amounts of DNA template are used.

The presence of microvariant alleles (alleles differing from one another by lengths other than the repeat length) complicates interpretation and assignment of alleles. There appears to be a correlation between a high degree of polymorphism, a tendency for microvariants and increased mutation rate (31,32).

9.B. DNA Extraction and Quantitation Methods and Automation Support

The DNA IQ™ System (Cat.# DC6700) is a DNA isolation system designed specifically for forensic and paternity samples (33). This novel system uses paramagnetic particles to prepare clean samples for STR analysis easily and efficiently and can be used to extract DNA from stains or liquid samples, such as blood or solutions. The DNA IQ™ Resin eliminates PCR inhibitors and contaminants frequently encountered in casework samples. With larger samples, the DNA IQ™ System delivers a consistent amount of total DNA. The system has been used to isolate and quantify DNA from routine sample types including buccal swabs, stains on FTA® paper and liquid blood. Additionally, DNA has been isolated from casework samples such as tissue, differentially separated sexual assault samples and stains on support materials. The DNA IQ™ System has been tested with the PowerPlex® Systems to ensure a streamlined process. See Section 9.F for ordering information.

For applications requiring human-specific DNA quantification, the Plexor® HY System (Cat.# DC1000) has been developed (34). See Section 9.F for ordering information.

For information about automation of Promega chemistries on automated workstations using Identity Automation™ solutions, contact your local Promega Branch Office or Distributor (contact information available at: **www.promega.com/support/worldwide-contacts/**), e-mail: **genetic@promega.com** or visit: **www.promega.com/idautomation/**

9.C. The Internal Lane Standard 600

The Internal Lane Standard 600 contains 22 DNA fragments of 60, 80, 100, 120, 140, 160, 180, 200, 225, 250, 275, 300, 325, 350, 375, 400, 425, 450, 475, 500, 550 and 600 bases in length (Figure 13). Each fragment is labeled with carboxy-X-rhodamine (CXR) and may be detected separately (as a fourth color) in the presence of PowerPlex® Y-amplified material. The ILS 600 is designed for use in each gel lane or CE injection to increase precision in analyses when using the PowerPlex® Y System. Protocols for preparation and use of this internal lane standard are provided in Section 5.

Note: The PowerPlex® Y System requires detection and definition of the 375-base peak of the ILS 600 to accurately size the largest alleles that may be observed.

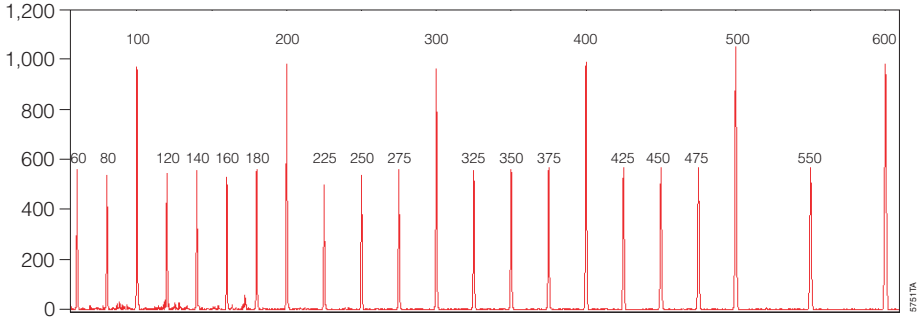

Figure 13. Internal Lane Standard 600. An electropherogram showing the Internal Lane Standard 600 fragments.

9.D. Preparing the PowerPlex® Y System PCR Amplification Mix

A worksheet to calculate the required amount of each PCR amplification mix component is provided in Table 6. Multiply the volume (µl) per reaction by the total number of reactions to obtain the final PCR amplification mix volume (µl).

Table 6. PCR Amplification Mix for the PowerPlex® Y System.

PCR Amplification Mix Component	Volume Per Reaction	×	Number of Reactions	=	Final Volume (µl)
Gold ST★R 10X Buffer	2.5µl	×		=	
PowerPlex® Y 10X Primer Pair Mix	2.5µl	×		=	
AmpliTaq Gold® DNA polymerase[1]	0.55µl (2.75u)	×		=	
nuclease-free water[2]	µl	×		=	
Per tube		×		=	
template DNA volume[2] (0.25–1ng)	up to 19.45µl	×		=	
total reaction volume	25µl	×		=	

[1]Assumes the AmpliTaq Gold® DNA polymerase is at 5u/µl. If the enzyme concentration is different, the volume of enzyme must be adjusted accordingly.

[2]The amplification mix volume and template DNA volume should total 25µl. Consider the volume of template DNA, and add nuclease-free water to the amplification mix to bring the final volume of the final reaction to 25µl.

9.E. Composition of Buffers and Solutions

10% ammonium persulfate

Add 0.05g of ammonium persulfate to 500µl of deionized water.

Blue Dextran Loading Solution

88.25%	formamide
15mg/ml	blue dextran
4.1mM	EDTA (pH 8.0)

Gold ST★R 10X Buffer

500mM	KCl
100mM	Tris-HCl (pH 8.3 at 25°C)
15mM	$MgCl_2$
1%	Triton® X-100
2mM	each dNTP
1.6mg/ml	BSA

TBE 10X buffer

107.8g	Tris base
7.44g	EDTA ($Na_2EDTA \cdot 2H_2O$)
~55.0g	boric acid

Dissolve Tris base and EDTA in 800ml of deionized water. Slowly add the boric acid, and monitor the pH until the desired pH of 8.3 is obtained. Bring the final volume to 1 liter with deionized water.

TE^{-4} buffer (10mM Tris-HCl, 0.1mM EDTA [pH 8.0])

1.21g	Tris base
0.037g	EDTA ($Na_2EDTA \cdot 2H_2O$)

Dissolve Tris base and EDTA in 900ml of deionized water. Adjust to pH 8.0 with HCl. Bring the final volume to 1 liter with deionized water.

TE^{-4} buffer with 20µg/ml glycogen

1.21g	Tris base
0.037g	EDTA ($Na_2EDTA \cdot 2H_2O$)
20µg/ml	glycogen

Dissolve Tris base and EDTA in 900ml of deionized water. Adjust to pH 8.0 with HCl. Add glycogen. Bring the final volume to 1 liter with deionized water.

9.F. Related Products

Fluorescent STR Multiplex Systems

Product	Size	Cat.#
PowerPlex® 16 System	100 reactions	DC6531
	400 reactions	DC6530
PowerPlex® 21 System	200 reactions	DC8902
PowerPlex® 18D System	200 reactions	DC1802
	800 reactions	DC1808
PowerPlex® 16 HS System	100 reactions	DC2101
	400 reactions	DC2100

Not for Medical Diagnostic Use.

Fluorescent STR Multiplex Systems (continued)

Product	Size	Cat.#
PowerPlex® ESX 16 System	100 reactions	DC6711
	400 reactions	DC6710
PowerPlex® ESX 17 System	100 reactions	DC6721
	400 reactions	DC6720
PowerPlex® ESI 16 System	100 reactions	DC6771
	400 reactions	DC6770
PowerPlex® ESI 17 Pro System	100 reactions	DC7781
	400 reactions	DC7780
PowerPlex® S5 System	100 reactions	DC6951
	400 reactions	DC6950

Not for Medical Diagnostic Use.

Accessory Components

Product	Size	Cat.#
PowerPlex® Matrix Standards, 310*	50µl (each dye)	DG4640
PowerPlex® Matrix Standards, 3100/3130*	25µl (each dye)	DG4650
2800M Control DNA*	25µl	DD7101
	500µl	DD7251
9948 Male DNA*	250ng	DD2061
9947A DNA*	250ng	DD1001
Internal Lane Standard 600	150µl	DG1071
Gold ST★R 10X Buffer	1.2ml	DM2411
Mineral Oil	12ml	DY1151
Water, Amplification Grade	6,250µl (5 × 1,250µl)	DW0991

*Not for Medical Diagnostic Use.

Sample Preparation Systems

Product	Size	Cat.#
DNA IQ™ System	100 reactions	DC6701
	400 reactions	DC6700
Casework Extraction Kit*	100 reactions	DC6745
Differex™ System*	50 samples	DC6801
	200 samples	DC6800
Maxwell® 16 Forensic Instrument*	1 each	AS3060
DNA IQ™ Reference Sample Kit for Maxwell® 16**	48 preps	AS1040
DNA IQ™ Casework Pro Kit for Maxwell® 16*	48 preps	AS1240
Plexor® HY System*	800 reactions	DC1000
	200 reactions	DC1001
Slicprep™ 96 Device	10 pack	V1391

*Not for Medical Diagnostic Use.
**For Research Use Only. Not for use in diagnostic procedures.

Polyacrylamide Gel Electrophoresis Reagents

Product	Size	Cat.#
Ammonium Persulfate, Molecular Grade	25g	V3131
TBE Buffer, 10X, Molecular Biology Grade	1L	V4251
Urea	1kg	V3171
Blue Dextran Loading Solution	3ml	DV4351

ART® Aerosol-Resistant Tips

Product	Volume	Size (tips/pack)	Cat.#
ART® 10 Ultramicro Pipet Tip	0.5–10µl	960	DY1051
ART® 20E Ultramicro Pipet Tip	0.5–10µl	960	DY1061
ART® 20P Pipet Tip	20µl	960	DY1071
ART® GEL Gel Loading Pipet Tip	100µl	960	DY1081
ART® 100 Pipet Tip	100µl	960	DY1101
ART® 100E Pipet Tip	100µl	960	DY1111
ART® 200 Pipet Tip	200µl	960	DY1121
ART® 1000E Pipet Tip	1,000µl	800	DY1131

[a]STR loci are the subject of U.S. Pat. No. RE 37,984, German Pat. No. DE 38 34 636 C2 and other patents issued to the Max-Planck-Gesellschaft zur Förderung der Wissenschaften, e.V., Germany.

[b]The purchase of this product does not convey a license to use AmpliTaq Gold® DNA polymerase. You should purchase AmpliTaq Gold® DNA polymerase licensed for the forensic and human identity field directly from your authorized enzyme supplier.

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